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Article

Screening an In-House Isoquinoline Alkaloids Library for New Blockers of Voltage-Gated Na⁺ Channels Using Voltage Sensor Fluorescent Probes: Hits and Biases

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Abstract: Voltage-gated Na⁺ (Na_V) channels are significant therapeutic targets for the treatment of cardiac and neurological disorders, thus promoting the search for novel Na_V channel ligands. With the objective of discovering new blockers of Na_V channel ligands, we screened an In-House vegetal alkaloid library using fluorescence cell-based assays. We screened 62 isoquinoline alkaloids (IA) for their ability to decrease the FRET signal of voltage sensor probes (VSP), which were induced by the activation of Na_V channels with batrachotoxin (BTX) in GH3b6 cells. This led to the selection of five IA: liriodenine, oxostephanine, thalamiculine, protopine, and bebeerine, inhibiting the BTX-induced VSP signal with micromolar IC₅₀. These five alkaloids were then assayed using the Na⁺ fluorescent probe ANG-2 and the patch-clamp technique. Only oxostephanine and liriodenine were able to inhibit the BTX-induced ANG-2 signal in HEK293-hNav1.3 cells. Indeed, liriodenine and oxostephanine decreased the effects of BTX on Na⁺ currents elicited by the hNav1.3 channel, suggesting that conformation change induced by BTX binding could induce a bias in fluorescent assays. However, among the five IA selected in the VSP assay, only bebeerine exhibited strong inhibitory effects against Na⁺ currents elicited by the hNav1.2 and hNav1.6 channels, with IC₅₀ values below 10 μM. So far, bebeerine is the first BBIQ to have been reported to block Na_V channels, with promising therapeutical applications.

Keywords: voltage-gated sodium channel; isoquinoline alkaloids; oxoaporphine; voltage sensor probes; Na⁺ fluorescent probe ANG-2; GH3b6 cells

1. Introduction

Voltage-gated sodium channels (Na_v channels) are heteromultimeric membrane proteins that play a major role in neuronal excitability and the excitation–contraction coupling of myocytes [1]. They are composed of a large pore-forming α -subunit encoded by nine genes (*scn1a*, *2a*, *3a*, *4a*, *5a*, *8a*, *9a*, *10a*, and *11a*), thus giving a total of nine Na_v channel subtypes (Na_v1.1–Na_v1.9) [2]. Mutations leading to the gain of function of Na_v channels are responsible for a wide variety of pathologies, including epilepsy, chronic pain, and cardiac arrhythmia [3]. Therefore, searching for new Na_v channel modulatory drugs remain highly attractive for clinical and pharmaceutical purposes. In addition, Na_v channels represent a therapeutically validated target for five categories of inhibitory drugs: (i) anti-arrhythmics, (ii) anti-convulsants, (iii) antiepileptics, (iv) anesthetics, and (v) analgesics [4].

Alkaloids constitute a large and heterogeneous group of basic, heterocyclic, nitrogen-containing, natural compounds with a tremendous number of chemical structures [5]. They are mainly produced by plants, but they can also be found in bacteria, fungi, and animals. Although they are generally toxic, they also exhibit therapeutical properties with multi-target activities [6,7]. More than 27,000 structures of alkaloids have been described, and several have been used in traditional and conventional medicine, such as in analgesic (codeine, morphine), anti-tumoral (taxol), anti-arrhythmic (quinidine), antispasmodic (atropine, hyoscyamine), muscle relaxant (tubocurarine), and antihypertensive (canescine, rescinamine) drugs [6,7]. Despite having been studied for decades, the high structural diversity of alkaloids with regard to their pharmacological properties makes them a highly relevant chemical group for drug screening assays.

Eight distinct Na_v channel binding sites (1 to 8) have been described for natural ligands [4,8]. Historically, tetrodotoxin (TTX) and saxitoxin (STX), two hydrophilic microbial neurotoxins that are structurally related to guanidine alkaloids, were the first to be characterized as potent inhibitors of Na_v channels through their binding to Site 1, localized on the external side of the ion channel pore [4,9,10]. Since then, Na_v channel subtypes have been classified as TTX-sensitive (TTX-S) or TTX-resistant (TTX-R) because they are blocked by nanomolar and micromolar concentrations, respectively [11,12].

Other alkaloids with various structures, mainly of vegetal origin, target Na_v channels (Figure 1). Lipid-soluble alkaloid toxins, including batrachotoxin (BTX) and veratridine (VTD) as well as aconitine bind to Site 2 [4,8]. These toxins bind to open Na_v channels and maintain them in this configuration, thus preventing their transition to the inactivation state [8,13,14]. BTX, VTD, and aconitine are considered as Na_v channel activators, which lead to the membrane depolarization of excitable cells and their firing [8]. BTX binds to Na_v channels with very high affinity, behaving as a full activator, whereas VTD and aconitine behave as partial activators [8].

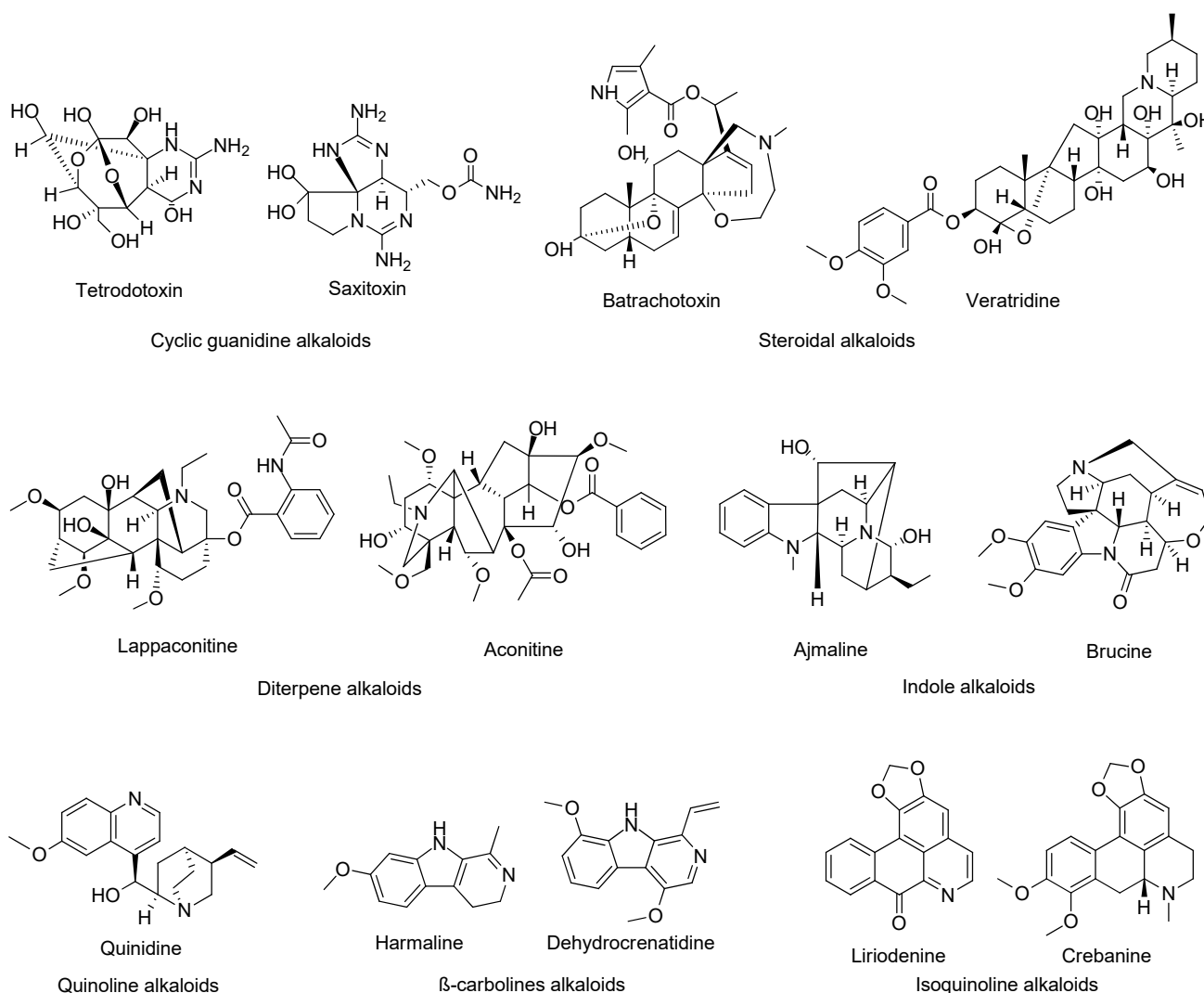


Figure 1. Structural diversity of natural alkaloids active on Na_v channels. Tetrodotoxin and saxitoxin are two marine guanidine alkaloids. Veratridine, aconitine, and batrachotoxin are activators of the Na_v channels. Lappaconitine, quinidine, and ajmaline are two inhibitors used as clinical drugs. Liriodenine and crebanine, two IA, block the Na_v channels. Lappaconitine, which is structurally related to aconitine [15], exhibits potent analgesic properties [16] and has recently been proposed for the treatment of chronic pain [17]. Ajmaline, isolated from *Rauwolfia serpentina*, inhibits the Na_v1.5 channels [18–21]. Ajmaline is used in the diagnosis and treatment of cardiac arrhythmia [22]. Brucine, extracted from *Nux vomica*, exhibits antinociceptive properties mediated by Na_v channel inhibition [23]. Harmaline from *Peganum harmala* [24] and dehydrocrenatidine from *Picrasma quassioides* are used to treat pain [25].

Isoquinoline alkaloids (IA) represent one of the most diverse and abundant groups of plant compounds. They are biosynthesized from tyrosine, leading to several chemical subgroups such as benzylisoquinoline, aporphine, protoberberine, protopine, morphinan, and many others [7,26]. Until now, only two of the large aporphine alkaloid subgroups (>1000 members) have been described as Na_v channel blockers: liriodenine from *Fissistigma glaucescens* [27] and crebanine from *Stephania* sp. [28] (Figure 1). Both compounds exhibit anti-arrhythmic activities mediated by the inhibition of multiple cardiac ion channels, such as the Na_v, Ca_v, and K_v channels [27–29]. However, this IA group has been poorly studied for their effects on Na_v channels. Therefore, in this work, we aimed to identify new Na_v channel blockers among IA from an in-house library composed of plant alkaloids.

We focused our studies on the IA subgroups aporphine, 1-benzyl isoquinoline (BIQ), protoberberine, and bisbenzylisoquinoline (BBIQ) in our search for new blockers of Na_V channels. The screening assay consists in the measurement of the FRET voltage sensor probe (VSP) signal elicited by the activation of Na_V channels that are endogenously expressed in GH3b6 cells by BTX in the absence and in the presence of 62 IA from our in-house natural compounds library. This allowed us to identify five putative Na_V channel blockers: liriodenine, oxostephanine, thalmituline, protopine, and bebeerine. These five IA were assayed for their ability to block the ANG-2 fluorescence emission induced by BTX in HEK293-expressing hNav1.3 and then to inhibit Na^+ currents elicited by the hNav1.2, hNav1.3, and hNav1.6 channels via automated patch-clamp electrophysiology. Our findings highlighted that the use of the activators of Na_V channels, such as BTX, which exhibits a “foot-in-the-door” mechanism and thus freezes the Na_V channels in an open state, could compromise the discovery of selective and potent inhibitors.

2. Results

2.1. VSP-FRET Assays Using GH3b6 for Pharmacological Characterization of Na_V Channels

The GH3b6 cell line was selected for its wide use as a cellular model for pharmacological studies on Na_V channels [30–35]. Here, we used RT-qPCR to establish the Na_V channel gene expression profile of GH3b6 cells. Our data showed the significant expression of four Nav channel transcripts (*scn1a*, *scn2a*, *scn3a*, and *scn8a*), which encoded four neuronal and TTX-sensitive Na_V channel subtypes (Nav1.1, 1.2, 1.3, and 1.6, respectively) (Figure 2). The rank order of the expression levels of these transcripts was *scn3a* > *scn8a* > *scn2a* > *scn1a* ($p < 0.0001$). The amplification of *scn3a* was observed with the lowest Ct-values (Ct = 22.9, for 10 ng of ADNc). Our data indicate that GH3b6 cells mainly expressed the neuronal TTX-sensitive Na_V 1.3 channel subtype, which thereby tends to be consistent with earlier observations [35–37].

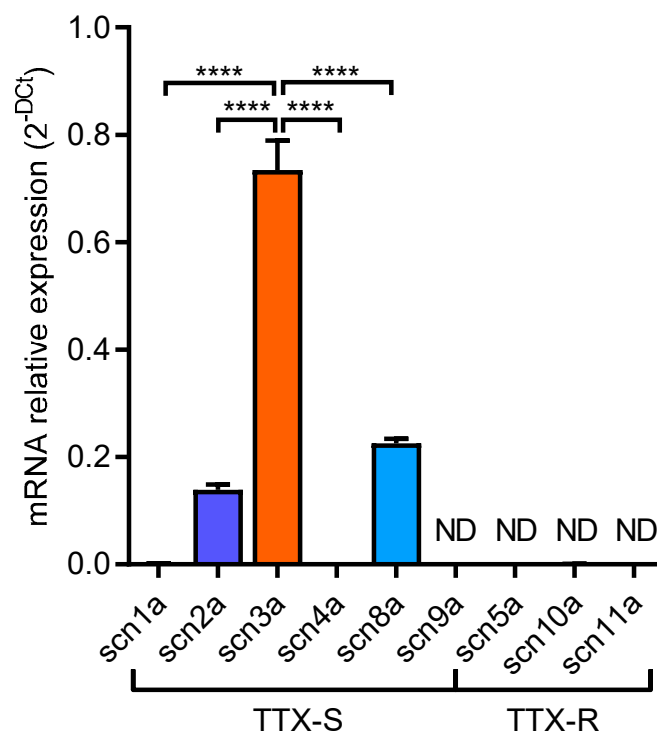


Figure 2. GH3b6 cells express TTX-S Nav channels. Nav channel gene expression profile of GH3b6 cells. The histogram illustrates the expression repertoire of Nav channel genes in GH3b6 cells as established by RT-qPCR. Data are the mean of mRNA relative expression $\pm$ SEM ($n = 6$). ND: Not Detected. Significance tests between groups of data were performed using one-way ANOVA followed by Bonferroni’s multiple comparison test (**** $p < 0.0001$).

Next, we measured the fluorescent resonance transfer (FRET) between CC2-DMPE and DisBAC2(3) in GH3b6 cells to characterize the change in the membrane potential induced by BTX as it is considered as a full activator of Na_v channels [8,13,14]. The fluorescence emission of CC2-DMPE increased, which is in parallel with the decrease of DisBAC2(3) fluorescence in the presence of increasing concentrations of BTX (from 0.01 to 3 μM) (Figure 3a). Subsequently, the VSP fluorescence emission ratio increased in a concentration-dependent manner. The data were best fitted by the Hill–Langmuir equation with a variable slope for a bimolecular reaction with an EC_{50} value of $0.27 \pm 0.06 \mu\text{M}$ (Figure 3b). At 100 nM, TTX fully abolished the VSP fluorescence emission ratio induced by 1 μM BTX (Figure 3b). The TTX inhibitory effects were fitted as described above, with an IC_{50} value of $8.58 \pm 1.67 \text{ nM}$ (Figure 3b). Indeed, BTX binds to the open state of Na_v channels and consequently prevents their closing in an irreversible manner. Then, the injection of Na^+ , which depolarized the membrane, led to the activation of the Na_v channels and allowed for BTX binding. Since this interaction is irreversible, the cells became leaky to Na^+ . This led to an Na^+ influx, which was counteracted by an Na^+ efflux mediated by Na^+/K^+ ATPase and other Na^+ transporters (e.g., NCX). This explains the slow kinetic of the fluorescence emission, which reached a plateau. This latter corresponds to an equilibrium between the Na^+ influx and the Na^+ efflux. Thus, the VSP signal induced by BTX was inhibited by TTX at a nanomolar concentration, in accordance with the presence of TTX-sensitive Na_v channels. Altogether, these results demonstrate that GH3b6 cells are suitable for a FRET-based VSP assay for the screening of Na_v channel inhibitors.

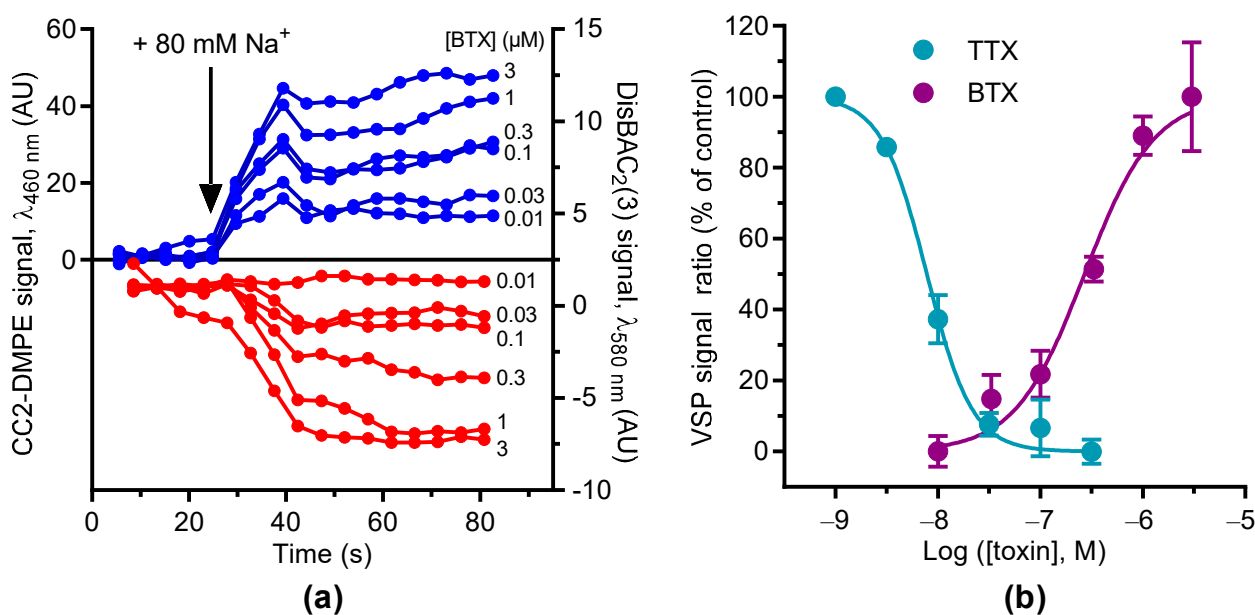


Figure 3. Monitoring the BTX activation of Na_v channels using VSP in GH3b6 cells. (a) Representative kinetic traces illustrating the fluorescence emission of CC2-DMPE ($\lambda = 460 \text{ nm}$, blue line) and DisBAC2(3) ($\lambda = 580 \text{ nm}$, red line) in the presence of increasing concentrations of BTX (from 0.01 to 3 μM) before and after raising the Na^+ concentration to 80 mM. Data are the mean of three wells. (b) Concentration–response curves of a BTX-induced VSP signal. BTX induced an increase in VSP signal ratio in a concentration-dependent manner. TTX decreased the VSP signal ratio induced by BTX at 1 μM in a concentration-dependent manner. Normalized data were calculated with four end points of the fluorescence emission ratio (Signal ratio CC2-DMPE/DisBAC2(3)) kinetics. Data were best fitted with the Hill–Langmuir equation with a variable slope. For BTX, $\text{EC}_{50} = 0.27 \pm 0.06 \mu\text{M}$, Hill slope = 1.21 ± 0.26 . For TTX, $\text{IC}_{50} = 8.58 \pm 1.67 \text{ nM}$, Hill slope = 1.53 ± 0.41 . Data are mean $\pm$ SEM ($n = 3$).

2.2. Screening Vegetal Alkaloids for Blockers of Na_V Channels Using VSP in GH3b6 Cells

Sixty-two IA (from IA1 to IA77) from our in-house library were chosen for their structure, none of which, except for liriodenine, had been described as a Na_V channel ligand (see supplementary Table S1, for trivial name, chemical class (CAS number), structure, plant origin, and reference). Thus, we first blind-tested these 62 IA for their ability to inhibit a BTX-induced VSP signal in GH3b6 cells (Figure 4). Figure 4a shows the BTX-induced VSP signal in the absence and in the presence of each IA. Thus, thirty samples exhibited an inhibition rate of 50% and more (Figure 4b). To select the potent inhibitors of Na_V channels, we used a threshold of 75% signal inhibition since the IA had been screened at rather high concentrations (10–50 $\mu\text{g}/\text{mL}$, corresponding to 12–20 μM) (Figure 4b). Then, ten IA were selected for further chemical analysis: (1) HPLC monitoring at two different wavelengths (210 nm and 280 nm) to evaluate their purity in %, (2) NMR structure validation, and (3) if necessary, mass spectrometry (MS) analysis and/or optical rotation to confirm IA identity (Table 1). It was only at this stage that the different samples were identified and the blind evaluation was stopped. Samples IA50 (tetrandrine), IA31 (tiliacorine), IA36 (sukhodanin), and IA42 (gyrocarpusine) were rejected because of their low purity (<75% at 210 nm and 280 nm). Therefore, we retained the following five IA for further pharmacological characterization: oxostephanine (IA14 and IA24), liriodenine (IA39), thalamiculine (IA49), bebeerine (IA52), and protopine (IA69) (Table 1).

The NMR spectra of IA24, 14, 39, 49, 52, and 69 confirmed their structures (Supplementary Figures S1–S4). Sometimes, MS and $[\alpha]_D^{20^\circ\text{C}}$ were necessary. Interestingly, oxostephanine and liriodenine are two members of the small subgroup of oxoaporphines.

Oxostephanine, liriodenine, thalamiculine, bebeerine, and protopine inhibited the BTX-induced VSP signal in GH3b6 cells in a concentration-dependent manner, with IC_{50} values in the micromolar range (Figure 5). Oxostephanine and liriodenine were able to abolish the BTX-induced VSP signal with similar IC_{50} values (5–6 μM). In contrast, thalamiculine, bebeerine, and protopine decreased BTX response up to 31–65%, but with lower IC_{50} values (0.4–4 μM) (Table 2). The Hill coefficient values were close to 1 for these five IA, suggesting a bimolecular interaction between them and their molecular targets.

2.3. Effects of Oxostephanine, Liriodenine, Thalamiculine, Bebeerine, and Protopine on Intracellular Na^+ Concentration in a Stable Cell Line Expressing $\text{hNa}_V1.3$

To further characterize the inhibitory effects of liriodenine, oxostephanine, thalamiculine, protopine, and bebeerine on Na_V channels, these IA were assayed on a clonal HEK293 cell line expressing the human $\text{Na}_V1.3$ channel subtype (HEK293- $\text{hNa}_V1.3$) (Figure 6). We used the ANG-2 Na^+ probe, which has been proven to be the most reliable in monitoring the Na^+ intracellular concentration ($[\text{Na}^+]_i$) [38]. At 3 μM , BTX induced a slow increase of the ANG-2 fluorescence ratio, which was totally blocked by 1 μM TTX ($92 \pm 5\%$, $p < 0.0001$, Figure 6a,b). Oxostephanine, liriodenine, and bebeerine inhibited the BTX-induced $[\text{Na}^+]_i$ elevation in HEK293- $\text{hNa}_V1.3$ up to $43 \pm 12\%$ ($p < 0.01$), $50 \pm 10\%$ ($p < 0.001$), and $18 \pm 2\%$ ($p < 0.0001$) (Figure 6a,b). However, no significant inhibitory effects on $\text{hNa}_V1.3$ channels were observed with thalamiculine ($p = 0.08$) and protopine ($p = 0.15$) (Figure 6b). Altogether, these results partially agreed with data obtained in VSP assays, showing that only oxostephanine, liriodenine, and bebeerine could be considered as putative inhibitors of Na_V channels.

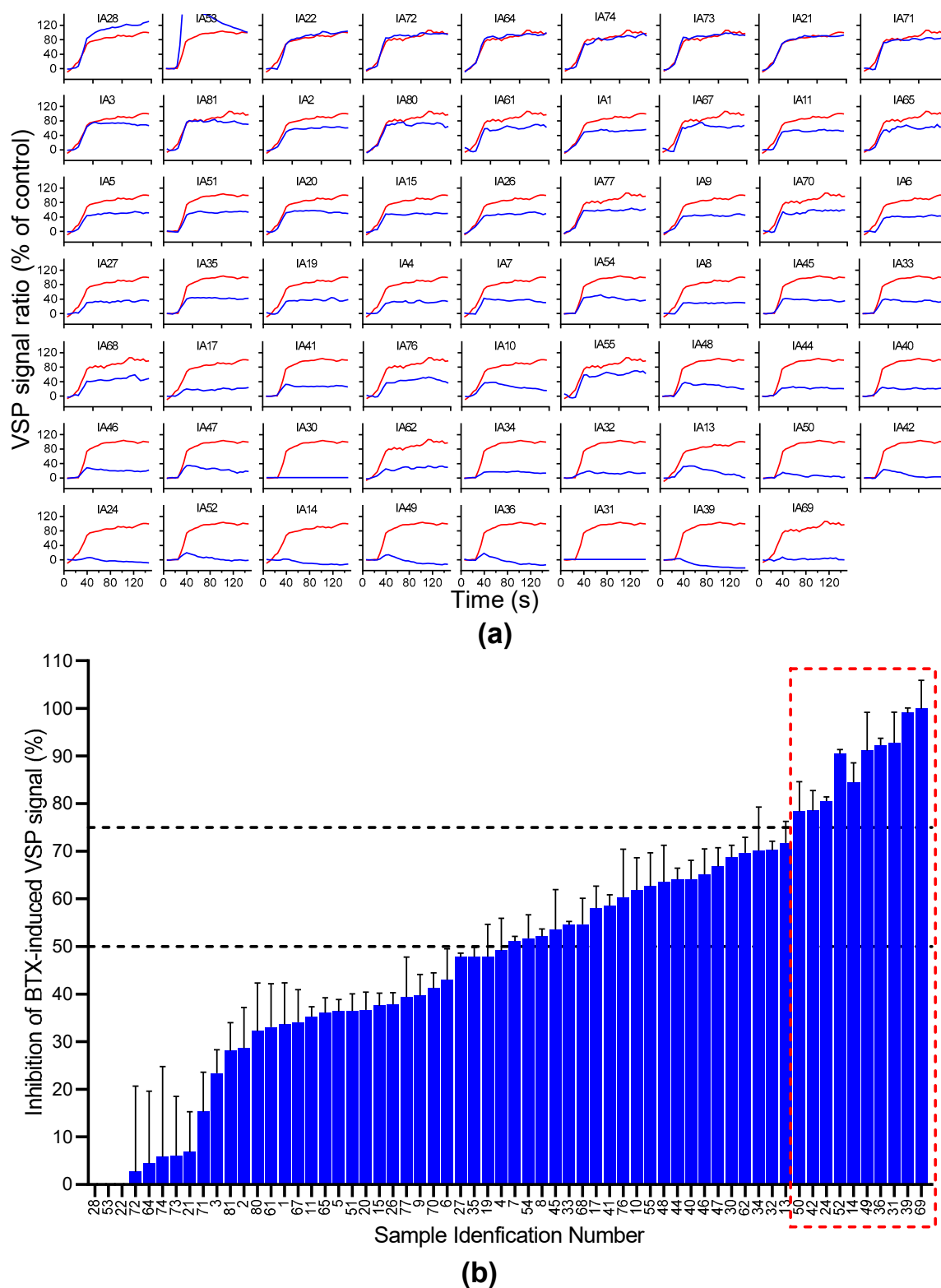


Figure 4. Screening of 62 samples from an in-house library of plant alkaloids for inhibitors of Nav channels. (a) Average kinetic ratios of VSP signal obtained in the presence of BTX alone (red line) and BTX co-incubated with IA (blue line), before and after injection of the depolarizing solution (at 30 s). Red and blue traces correspond to distinct experiments. x axis: fluorescence ratio (a.u.), y axis: time (s). (b) The histogram bars correspond to the inhibition of the BTX-induced VSP signal (in %). Ten IA induced at least 75% of the inhibition of BTX-induced depolarization (red dashed rectangle). Data are mean $\pm$ SEM ($n = 3$).

Table 1. Chemical analysis of IA selected for their ability to inhibit a BTX-induced VSP signal.

Sample	Inhibitory Effects (%)	Alkaloid Name	Plant Name	210 nm * (%)	280 nm * (%)	¹ HNMR, (MS, [α] _D ^{20°C})	Acceptance
IA50	78.3	tetrandine	<i>Pachygone dasycarpa</i> (Menispermaceae)	76	67	ND	No
IA42	78.5	gyrocarpusine	<i>Gyrocarpus americanus</i> (Hernandiaceae)	68	52	ND	No
IA24	80.5	oxostephanine	<i>Stephania venosa</i> (Menispermaceae)	77	98	Confirmed	Yes
IA52	81.9	bebeerine	<i>Curarea candicans</i> (Menispermaceae)	83	100	Confirmed	Yes
IA14	84.4	oxostephanine	<i>Stephania venosa</i> (Menispermaceae)	91	99	Confirmed	Yes
IA49	91.1	thalmiculine	<i>Thalictrum cultratum</i> (Ranunculaceae)	93	85	Confirmed	Yes
IA36	92.2	sukhodanin	<i>Stephania venosa</i> (Menispermaceae)	65	81	ND	No
IA31	92.8	tiliacorinine	<i>Tiliacora racemosa</i> (Menispermaceae)	73	81	ND	No
IA39	99.1	liriodenine	<i>Stephania venosa</i> (Menispermaceae)	77	99	Confirmed	Yes
IA69	99.9	protopine	<i>Corydalis majori</i> (Fumariaceae)	85	75	confirmed	Yes

This table shows the list of IA which inhibited a BTX-evoked VSP signal at more than 65% in GH3b6 cells. * Each compound was analyzed by HPLC monitored at two different wavelengths (210 and 280 nm) in order to evaluate their purity in %; below 75%, the compound was rejected. Then, the NMR-¹H (sometimes MS and [α]_D^{20°C}) was performed to confirm the IA structure. ND: not determined.

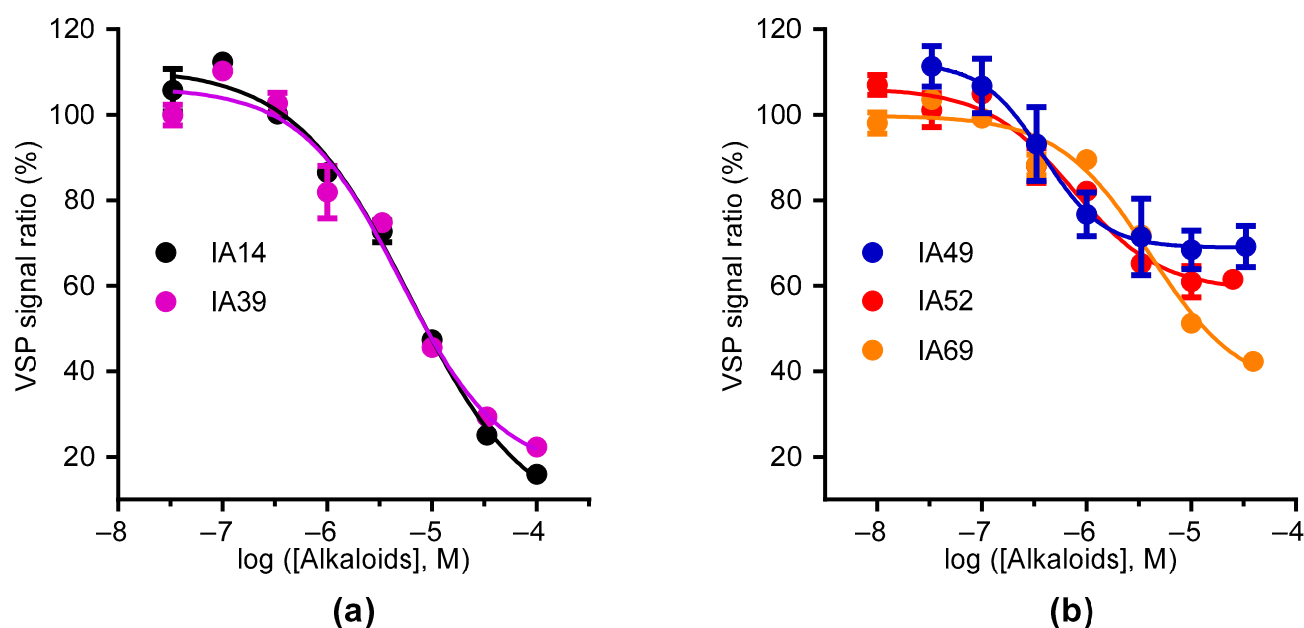


Figure 5. Concentration–inhibition curves of oxostephanine, liriodenine, thalmiculine, bebeerine, and protopine for the BTX-induced VSP FRET signal in GH3b6 cells. (a) Concentration–inhibition responses of the BTX-induced VSP signal to increasing concentrations of oxostephanine (IA14) and liriodenine (IA39). (b) Concentration–inhibition responses of the BTX-induced VSP signal to increasing concentrations of thalmiculine (IA49), bebeerine (IA52), and protopine (IA69). The data were best fitted by non-linear regression to the Langmuir–Hill equation with a variable slope and are shown in Table 2. Data are mean ($\pm$ SEM) and representative of two independent experiments ($n = 6$).

Table 2. Curve fitting parameters of the concentration–inhibition relationships of oxostephanine, liriodenine, thalamiculine, bebeerine, and protopine.

Sample	Alkaloid Name	Alkaloid Type	IC ₅₀ (μM)	Hill Slope	Maximum of Inhibition
IA14	oxostephanine	oxoaporphine	6.11 ± 0.09	0.78 ± 0.12	95.78 ± 6.99
IA39	liriodenine	oxoaporphine	5.00 ± 0.10	0.91 ± 0.19	83.78 ± 6.59
IA49	thalamiculine	BBIQ	0.38 ± 0.19	1.50 ± 0.93	31.07 ± 4.04
IA52	bebeerine	BBIQ	0.74 ± 0.11	1.04 ± 0.29	41.04 ± 3.44
IA69	protopine	protopine	3.97 ± 0.10	0.99 ± 0.19	64.67 ± 5.50

This table shows the IC₅₀, Hill coefficient, and the maximum of inhibition values determined from non-linear regression analysis of concentration–inhibition curves of BTX-evoked VSP signal by oxostephanine, liriodenine, thalamiculine, bebeerine, and protopine of Figure 5. Data are mean ± SEM (*n* = 6).

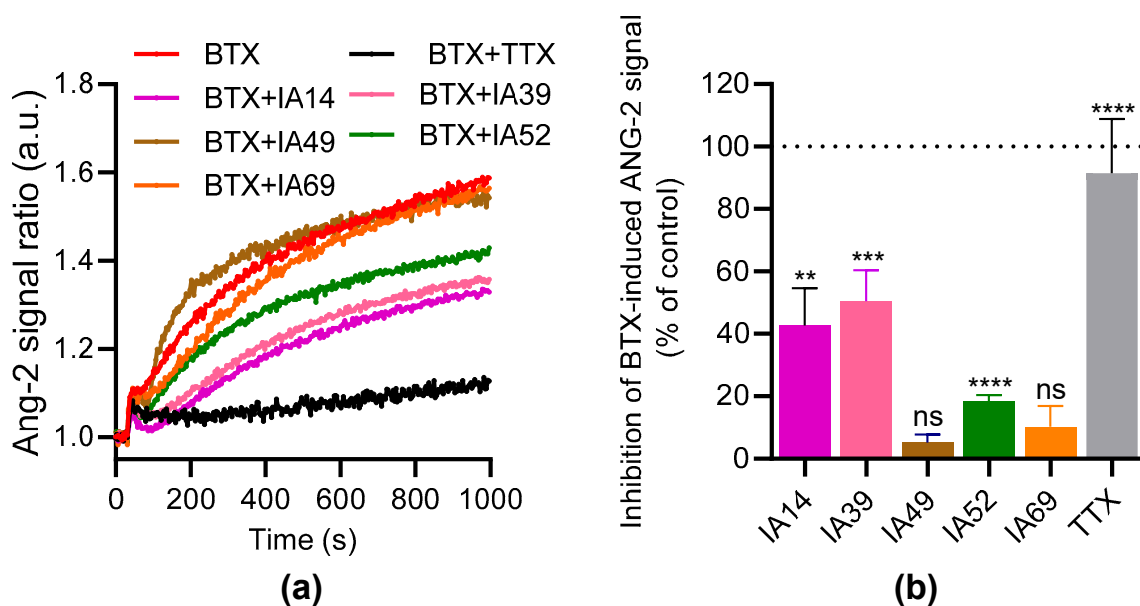


Figure 6. Effects of oxostephanine, liriodenine, thalamiculine, bebeerine, and protopine on the ANG-2 signal induced by BTX in HEK293-hNav_{1.3} cells. (a,b) Representative kinetic traces illustrating the fluorescence emission of ANG-2 probe induced by the injection of BTX (3 μM) without or with oxostephanine (IA14), liriodenine (IA39), bebeerine (IA52), thalamiculine (IA49), and protopine (IA69) at 10 μM each. TTX (1 μM) was used as control. (c,d) The histogram bars represent the inhibition rate (in %) of each IA at 10 μM on the ANG-2 fluorescence signal induced by BTX. Normalized data were calculated as described for the VSP signal. Data are mean ± SEM of 3–5 independent experiments. Significance was evaluated by two-tailed unpaired *t*-tests between each group, with zero as a hypothetical value. **, *p* < 0.01; ***, *p* < 0.001; ****, *p* < 0.0001; ns: not significant.

2.4. Characterization of the Effects of Oxostephanine, Liriodenine, Thalamiculine, Bebeerine, and Protopine on Na⁺ Currents Elicited by the hNav_{1.2}, hNav_{1.3}, and hNav_{1.6} Channels

To validate the inhibitory effects of oxostephanine, liriodenine, thalamiculine, bebeerine, and protopine on Na_V channels, automated patch-clamp experiments were performed using three different cell lines stably expressing the Na_V channel subtypes detected by RT-qPCR in GH3b6 cells. These five IA were assayed on the Na⁺ currents elicited by the depolarizing step in the three cell lines, which stably expressed the hNav_{1.2} (HEK293- hNav_{1.2} cells), hNav_{1.3} (HEK293- hNav_{1.3} cells), and hNav_{1.6} channels (HEK293- hNav_{1.6} cells). At 10 μM, oxostephanine, liriodenine, thalamiculine, and protopine had no effects on the hNav_{1.2}, hNav_{1.3}, and hNav_{1.6} channels (Figure 7a and Supplementary Figure S5). At the same concentration, bebeerine was able to inhibit the Na⁺ currents elicited by the hNav_{1.2}, hNav_{1.3}, and hNav_{1.6} channels (Figure 7a,b). The inhibition rate of bebeerine on these three Na_V channels did not depend on the V_m (two-way ANOVA test followed by Holm–Sidak’s multiple comparison test, *p* < 0.001). At a V_m between −20 mV and

30 mV, bebeerine had stronger inhibitory effects on the hNav1.3 (60–64%, $n = 38$) and hNav1.6 channels (70–74%, $n = 32$) than on the hNav1.2 channel (42–50%, $n = 38$, Figure 7c). Notably, bebeerine significantly inhibited the hNav1.6 channel to a higher extent ($74 \pm 3\%$, $n = 32$) than the hNav1.3 channel ($64 \pm 4\%$, $n = 48$; $p < 0.05$), at -20 mV and -15 mV, respectively (Figure 7c). Altogether, these data show that bebeerine efficiently blocked the hNav1.2, hNav1.3, and hNav1.6 channels, with the following rank order: hNav1.6 > hNav1.3 > hNav1.2.

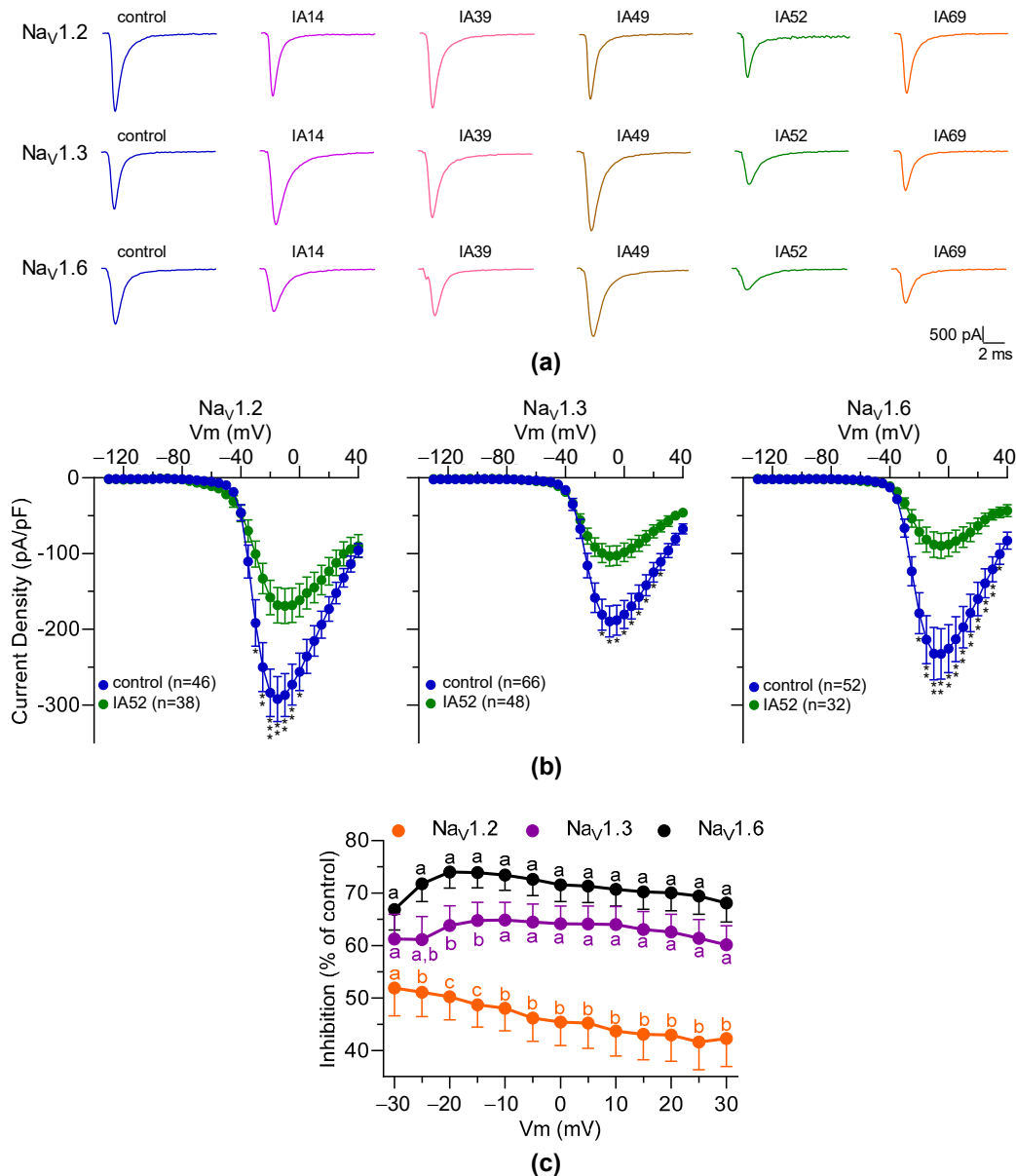


Figure 7. Effects of oxostephanine, liriodenine, thalamiculine, bebeerine, and protopine on Na^+ currents elicited by the hNav1.2, hNav1.3, and hNav1.6 channels. Oxostephanine (IA14), liriodenine (IA39), thalamiculine (IA49), bebeerine (IA52), and protopine (IA69) were assayed at $10 \mu\text{M}$ each on the hNav1.2, hNav1.3, and hNav1.6 channels stably expressed in HEK293T cells. Control experiments were performed with an extracellular solution containing 0.1% of DMSO corresponding to the vehicle. (a) Examples of Na^+ current traces elicited at $+20$ mV in the presence and in the absence of IA14, IA39, IA49, IA52, and IA69. (b) I-V relationship curves obtained by plotting the mean of peak current density to V_m . Data are shown as mean $\pm$ SEM. Multiple unpaired t -test corrected with the Holm–Sidak method was used to compare groups. *, $p < 0.05$; **, $p < 0.001$; ***, $p < 0.0001$. (c) Comparison of the inhibitory effects of IA52 on the hNav1.2, hNav1.3,

and hNa_V1.6 channels. The inhibition rate induced by IA52 on the Na⁺ currents was normalized ($V_m = -30$ to 30 mV). Data were plotted as mean $\pm$ SEM and analyzed for statistical significance using a two-way ANOVA test with Holm–Sidak’s multiple comparison test ($p < 0.0001$). Different letters (a, b, and c) above (hNa_V1.2 and hNa_V1.6) and below (hNa_V1.3) indicate significant differences between groups. p -values are shown in Supplementary Table S1.

Figure 8 illustrates the effects of bebeerine (at $10 \mu\text{M}$) on the voltage-dependent activation and inactivation of the hNa_V1.2, hNa_V1.3, and hNa_V1.6 channels. All fitting parameters are summarized in Table 3. There was no significant change in both the half-activation and inactivation potential ($V_{1/2}$) of the Na⁺ currents elicited by the hNa_V1.2 channels ($p = 0.72$ and $p = 0.69$, respectively) nor by the hNa_V1.6 channels ($p = 0.71$ and $p = 0.32$, respectively) in the presence of bebeerine (Figure 8a,c). Concerning the hNa_V1.3 channel, bebeerine induced a slight negative shift in $V_{1/2}$ activation (-3.6 ± 1.0 , $p < 0.001$), but not in $V_{1/2}$ inactivation ($p = 0.08$, Figure 8b). However, a significant increase of slope values for both the activation and inactivation curves was observed for each Na_V channel subtype in the presence of bebeerine (Table 3).

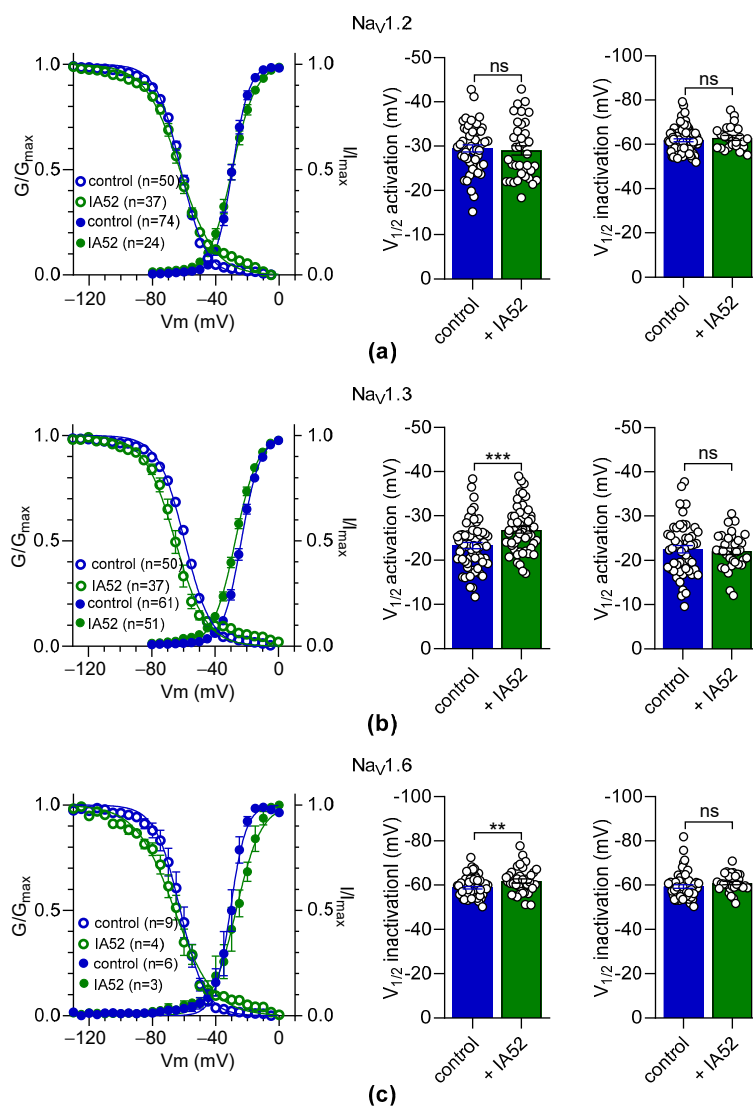


Figure 8. Effects of bebeerine on the activation and inactivation properties of Na⁺ currents elicited by the hNa_V1.2, hNa_V1.3, and hNa_V1.6 channels. The effects of bebeerine (IA52, $10 \mu\text{M}$) on the voltage-dependent activation and inactivation of the hNa_V1.2 (a), hNa_V1.3 (b), and hNa_V1.6 (c) channels were evaluated. Right panels (a–c): Activation and inactivation curves show data that were

fit with the Boltzmann equation. Middle and left panels: The scatter plots with bars show the $V_{1/2}$ of the activation and inactivation of the hNav1.2 (a), hNav1.3 (b), and hNav1.6 (c) channels. Control experiments were performed with extracellular solution containing 0.1% of DMSO corresponding to the vehicle. Data are shown as mean $\pm$ SEM. An unpaired *t*-test was used to analyze the significance between groups. ns: not significant; **, $p < 0.01$; ***, $p < 0.001$. Fitted parameters are shown in Table 3.

Table 3. Fitting parameters of the voltage-dependent activation and inactivation of Na⁺ currents elicited by the hNav1.2, hNav1.3, and hNav1.6 channels in the absence and in the presence of bebeerine.

Na _v Channel Subtype	Condition	Activation		Inactivation	
		$V_{1/2}$ ($\pm$ SEM, mV)	Slope ($\pm$ SEM)	$V_{1/2}$ ($\pm$ SEM, mV)	Slope ($\pm$ SEM)
hNav1.2	control	-29.5 ± 0.8 (n = 48)	3.9 ± 0.2	-61.8 ± 0.7 (n = 66)	-6.4 ± 0.1
	+bebeerine	-29.1 ± 1.1 (n = 36)	5.8 ± 0.3 ¹	-62.3 ± 1.1 (n = 21)	-8.5 ± 0.3 ⁴
hNav1.3	control	-23.3 ± 0.7 (n = 61)	4.9 ± 0.2	-58.2 ± 0.6 (n = 54)	-7.1 ± 0.2
	+bebeerine	-26.9 ± 0.7 (n = 51)	6.4 ± 0.3 ²	-60.2 ± 0.9 (n = 26)	-8.9 ± 0.3 ⁵
hNav1.6	control	-22.5 ± 0.8 (n = 52)	5.1 ± 0.2	-59.0 ± 0.8 (n = 50)	-7.7 ± 0.3
	+bebeerine	-22.1 ± 0.7 (n = 32)	7.0 ± 0.3 ³	-60.2 ± 0.7 (n = 24)	-9.0 ± 0.3 ⁶

This table shows the fitting parameters determined from the non-linear regression analysis of the voltage-dependent activation and inactivation of Na⁺ currents in the absence or presence of 10 μ M bebeerine (IA52, Figure 8). $V_{1/2}$, half potential (mV) for voltage-dependent activation or inactivation. Data are means $\pm$ SEM. Statistical significance between the control and test groups was evaluated with an unpaired *t*-test. ^{1,2,3,4,5}, $p < 0.0001$; ⁶, $p < 0.001$.

Oxostephanine and liriodenine did not exhibit significant inhibitory effects at 10 μ M on Na⁺ currents elicited by the Nav1.3 channels, while both IA were able to efficiently inhibit the BTX-induced ANG-2 fluorescence signals in HEK293-hNav1.3 cells. Thus, we assayed these two IA on the Na⁺ currents elicited by the hNav1.3 channel in the presence of 1 μ M BTX. As expected, BTX profoundly altered the Na⁺ currents elicited by the hNav1.3 channel by increasing the current densities and inhibition of inactivation (Figure 9a). Interestingly, in the presence of BTX, oxostephanine and liriodenine significantly decreased the current densities of the hNav1.3 channel at -60 mV (Figure 9a–c); however, only IA39 positively shifted the $V_{1/2}$ activation in the presence of BTX, but to a small extent ($p < 0.01$, Figure 9b).

We also examined the effects of oxostephanine and liriodenine on the inactivation process of hNav1.3 in the presence and in the absence of BTX. We evaluated the AUC of currents traces, which illustrate the inactivation rate of hNav1.3 channels. BTX induced the 55-fold increase of the AUC, indicating the decrease of the inactivation kinetic ($p < 0.0001$, data not shown). Interestingly, the slow-down of inactivation induced by BTX was significantly decreased by both IA14 and IA39 ($p < 0.05$, Figure 9b,c).

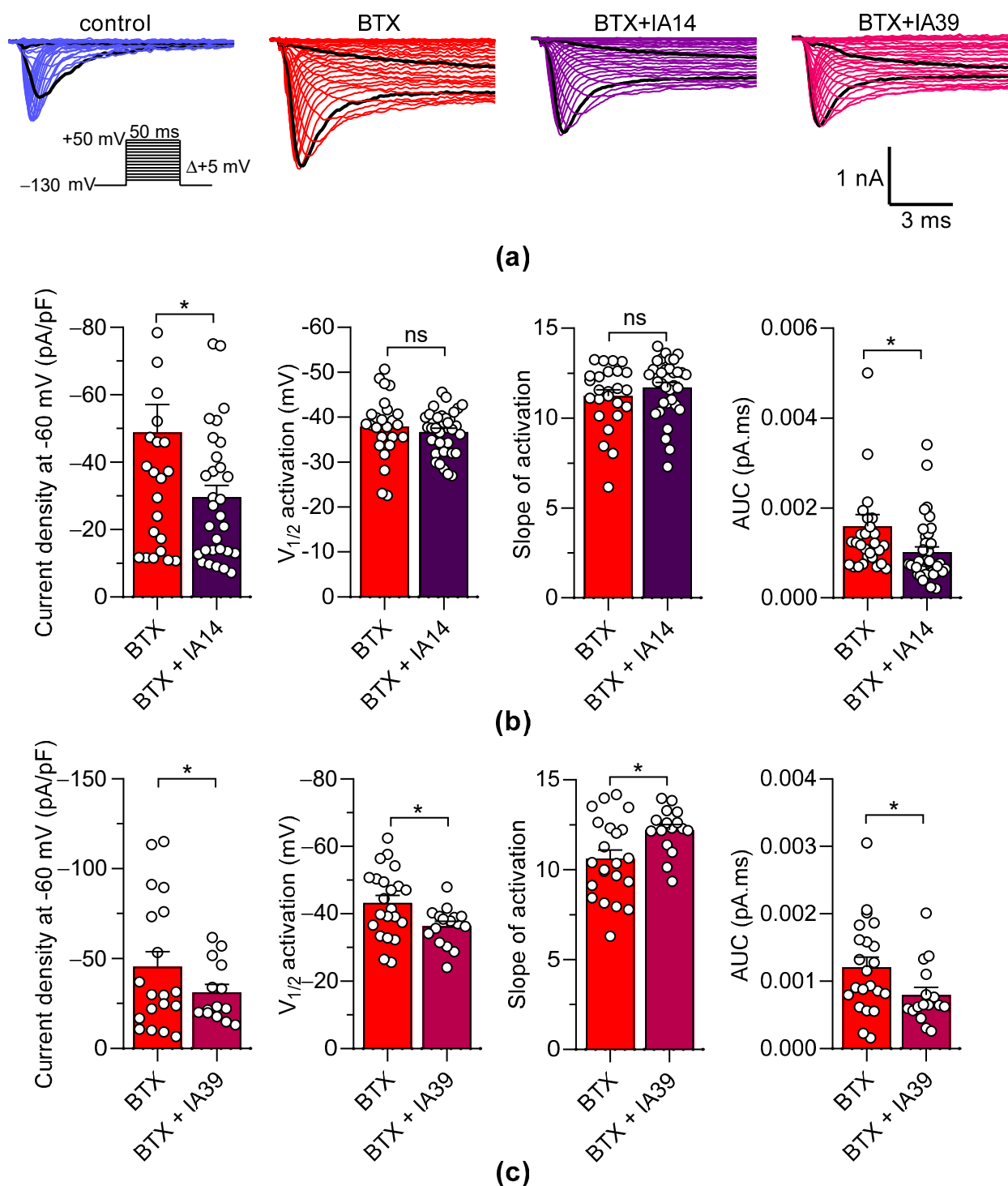


Figure 9. Effects of oxostephanine and liriodenine on Na⁺ currents elicited by hNav1.3 in the presence of batrachotoxin. Oxostephanine (IA14) and liriodenine (IA39) were assayed at 10 μM each on Na⁺ currents elicited by the hNav1.3 channel in the presence of batrachotoxin (BTX, 1 μM). (a) Representative examples of Na⁺ currents elicited by 50 ms depolarization steps (protocol inset). To illustrate the strong effect of 1 μM BTX, traces at −60 mV and 20 mV are shown in black. (b,c) The scatter plots with bars illustrate: the current densities at −60 mV, the $V_{1/2}$ of activation, the slope of activation curves, and the area under the curve (AUC in pA.ms) in the absence or in the presence of IA14 (b) and IA39 (c). Data are shown as mean ± SEM. Parametric or non-parametric statistical analyses were performed to determine the significance between groups. *, $p < 0.05$; ns: not significant.

3. Discussion

Our study aimed to identify new ligands of the Nav channels by screening vegetal IA from an in-house library. The IA family offers a large molecular diversity that has not been extensively studied for their activities against Nav channels. Using fluorescent VSP, we identified two oxoaporphine alkaloids (oxostephanine and liriodenine), two BBIQ (bebeerine and thalmiculine), and a protopine (protopine) which inhibited BTX-induced depolarization in GH3b6 cells, with IC₅₀ values in the micromolar range. However, by directly measuring [Na⁺]_i elevation, using the fluorescent probe ANG-2, only oxostephanine and liriodenine exhibited strong inhibitory effects against the hNav1.3 channel, which was stably expressed in HEK293 cells. Moreover, in patch-clamp experiments, only bebeerine was able to efficiently block the Na⁺ currents elicited by the hNav1.2 and hNav1.6 channels. We also showed that by altering the conformation of Nav channels, BTX allowed for the binding of oxostephanine and liriodenine.

The endocrine GH3b6 cell line has been extensively used as a cellular model to study Nav channels in patch-clamp experiments, notably those expressed in the central nervous system [39]. As we have previously demonstrated, these cells mainly express the endogenous Nav1.3 channel subtype [35]. Thus, these cells represent a good compromise in the search for or in characterizing new ligands of the Nav1.3 channel, a Nav channel involved in neuropathic pain [40]. To efficiently and rapidly screen our in-house IA library, we used VSP technology. Indeed, this strategy has been proven to be adapted for the detection and selection of ligands of Nav channels from chemical libraries or natural fractions [41–45]. For this strategy, it is necessary to activate Nav channels with selective activators, such as BTX or VTD. In our hands, fluorescent signals induced by BTX were stronger than those induced by VTD (data not shown), likely because BTX acts as a full activator [46]. However, despite an IC₅₀ value in the nanomolar range for TTX, we were very surprised that BTX-induced VSP signals were inhibited at more than 50%, with almost half of our in-house library in the range of 12–20 µM. Those higher concentrations could lead to a non-specific action on the cell that would modify the VSP signal indirectly. We then decided to select only IA with an inhibition rate of more than 75%, which yielded 10 hits. Among these hits and according to the physico-chemical analysis, five IA were selected: oxostephanine, liriodenine, thalmiculine, bebeerine, and protopine.

The VSP assays indicated that these five IA inhibit the BTX-induced VSP signals in GH3b6 cells with different efficiencies. While the oxoaporphines, oxostephanine, and liriodenine totally suppressed the BTX-induced VSP signals, thalmiculine, bebeerine, and protopine hardly inhibited these signals to 30–65%. This could be explained by the fact that BTX activates the Nav channel by binding to the pore with very high affinity, preventing any inactivation of the channels; this thereby promotes the identification of pore blockers, as it has already been discussed for VTD [47]. Thus, we first hypothesized that oxostephanine and liriodenine would behave as pore blockers, while bebeerine, thalmiculine, and protopine would exhibit a different mechanism of action. However, the use of VSP for the screening assay has been greatly debated, particularly in the search for selective Nav1.7 inhibitors [47,48]. Notably, this is because this technique requires the use of a neurotoxin activator, which can trap Nav channels in a particular conformation. Thus, this approach needs complementary techniques, such as patch-clamp recording or the use of fluorescent Na⁺ probes, both of which directly measure Nav channel activity. Here, we used ANG-2, which has been proven to be the most efficient in monitoring intracellular Na⁺ concentration [38].

Using this Na⁺ probe, we found that only oxostephanine and liriodenine were able to efficiently inhibit BTX-induced [Na⁺]_i elevations in hNav1.3-HEK293 cells. Indeed, at 10 µM, these two IA almost abolished BTX-induced [Na⁺]_i elevations, suggesting the micromolar affinities of both IA for the hNav1.3 channel in the presence of BTX. However, in our assay, thalmiculine, bebeerine, and protopine exhibited modest effects at 10 µM. Several hypotheses could be made: (1) thalmiculine, bebeerine, and protopine probably have weaker effects on the hNav1.3 channel than oxostephanine and liriodenine.

nine, (2) thalamiculine, bebeerine, and protopine could block other Na_V channel subtypes, (3) the spatio-temporal resolution of our assay did not allow us to highlight their blocking effects on the $\text{hNa}_V1.3$ channel, and (4) these three IA target other ion channels, such as the Cav and Kv channels, alleviating any change of $[\text{Na}^+]_i$ by the activation of NCX and other transporters. These data showed the advantages but also the limitations of using fluorescent probes in screening for Na_V channel ligands [38,47,49].

Our patch-clamp experiments showed that among the five IA selected by the VSP assays, only bebeerine behaved as a potent blocker of the $\text{hNa}_V1.2$ and $\text{hNa}_V1.6$ channels, with an IC_{50} value likely above $10\ \mu\text{M}$. Neither oxostephanine, liriodenine, thalamiculine, nor protopine had any inhibitory effects on the $\text{hNa}_V1.2$, $\text{hNa}_V1.3$, and $\text{hNa}_V1.6$ channels at $10\ \mu\text{M}$, revealing that they are likely to be low-potency blockers of these Na_V channel subtypes. We could not use higher concentrations of these compounds because of their low solubility and the non-specific effects of the DMSO concentrations. These unexpected results could be explained by the bias induced by BTX as a Na_V activator in fluorescent-based assays, as it has been previously discussed for VTD [47]. Just as VTD, BTX is known to bind within the pore, thus trapping Na_V channels in an open state [8,13,50], allowing for the binding of low-affinity blockers [47]. This assumption was confirmed by the fact that oxostephanine and liriodenine decreased the BTX effects on both the activation and inactivation properties of the $\text{hNa}_V1.3$ channel, while both IA had no effects on the inactivation properties of this Na_V channel in the absence of BTX. Thus, we assumed that both IA bind to Na_V channels in the presence of BTX, which traps Na_V channels in an open state. Altogether, our data highlighted that fluorescent-based screening assays on BTX-activated Na_V channels could compromise the finding of potent and selective blockers of these ion channels, and that the patch-clamp technique remains unavoidable for the accurate characterization of Na_V channel ligands.

Oxostephanine and liriodenine are two members of the oxoaporphine subgroup, and they only differ by a methoxy group on the benzyl part of the skeleton of this IA subgroup. Previous studies have demonstrated the ability of liriodenine to suppress Na^+ currents recorded in isolated cardiomyocytes with an IC_{50} value in the micromolar range [27,51], which is close to what we observed. Since cardiomyocytes mainly express $\text{Na}_V1.5$ [2], it is possible that liriodenine preferentially blocks this Na_V channel subtype. Several studies have shown that oxostephanine and liriodenine share some biological activities, such as antimicrobial and anti-cancer properties [52,53]. Liriodenine has been proven to target ion channels such as the K_V , Ca_V , and Na_V channels [27], but it also targets receptors such as the GABAA, nicotinic, and muscarinic receptors [54–56]. The high structural similarity between oxostephanine and liriodenine certainly supports the fact that they might share the same pharmacological targets, such as the Nav channels.

Finally, for the first time, we determined that bebeerine exhibits inhibitory effects on Na_V channels, with a higher potency on the $\text{hNa}_V1.6$ channel than on the $\text{hNa}_V1.2$ and $\text{hNa}_V1.3$ channels. Bebeerine is the enantiomer of curine, which has been extensively studied and previously described as an L-Type Ca_V channel blocker [57]. Thus, it is conceivable that bebeerine might also inhibit L-Type Ca_V channels. Interestingly, no BBIQ has been reported as a Na_V channel blocker so far, and since this IA subgroup is particularly large, further screening by an automated patch-clamp could be of interest. In addition, further studies are required to provide more information on the selectivity and also the blocking mechanism of bebeerine towards all Na_V channel subtypes.

4. Materials and Methods

4.1. Chemicals

A voltage sensor probe (VSP) set (CC2-DMPE and DisBAC₂(3)), VABSC-1 (Voltage Assay Background Suppression Compound), and Pluronic®-F127 were purchased from Invitrogen (Carlsbad, CA, USA). BTX and TTX were purchased from Latoxan (Valence, France). Asante NaTRIUM Green-2 (ANG-2) was purchased from Interchim (Montluçon,

France). All other reagents (Ponceau 4R) and solvents were obtained from Sigma-Aldrich (Saint-Louis, MO, USA) or Fisher Scientific (Waltham, MA, USA).

4.2. In-House Library of Vegetal Alkaloids

More than 300 alkaloids from plants have been collected in a library of the SONAS laboratory (EA 921, University of Angers) in the last four decades. These compounds were extracted and purified from at least 14 plant families (*Annonaceae*, *Berberidaceae*, *Fumariaceae*, *Lauraceae*, *Menispermaceae*, etc.), which were collected from all over the world (Bolivia, Colombia, France, Indonesia, Jordan, etc.) (Supplementary Figure S5). They were stored dried at room temperature. In a blind test, 62 IA associated with 10 different structural subtypes (Supplementary Table S2) were screened for their ability to inhibit BTX-evoked depolarization in GH3b6 cells. Each IA or fraction was solubilized in DMSO at 1–5 mg/mL, stored, and sheltered from light at 4 °C.

4.3. Cell Culture

GH3b6 cells are a subclone of the GH3 pituitary cell line [58] and were a generous gift from Dr. Françoise Macari (IGF, Montpellier, France). GH3b6 cells were cultivated from passage 14 until passage 32 at 37 °C/5% CO₂ in Dubelcco's Modified Eagle Medium (DMEM)/F12 (Lonza, Basel, Switzerland), supplemented with 10% FBS (Lonza, Basel, Switzerland), 1 mM L-glutamine, and 1 mM penicillin and streptomycin solution. For the fluorescence assay, the GH3b6 cells were seeded in black-walled, clear-bottom, 96-well, tissue culture-treated, and sterile imaging plates (Corning HazeBrouck, France), at a density of 10,000 cells per well, and incubated for 24 h before starting the experiment. The human Na_v1.3 (hNa_v1.3) channel-expressing cell line (HEK293) used in this work was a generous gift from Massimo Mantegazza (IPMC, Nice, France). HEK293-hNa_v1.3 cells were cultivated from passage 20 until passage 35 at 37 °C/5% CO₂ in a DMEM high glucose medium (4.5 g/l), supplemented with 10% FBS, pyruvate, L-glutamine, and penicillin and streptomycin solution, each one at 1 mM, completed with 800 µg/mL geneticin G418. For the fluorescence assays, the cells were plated at a density of 50,000 cells per well in black-walled, clear-bottom, 96-well plates and then cultured at 37 °C and 5% CO₂ for 24 h before the experiments. HEK293 stably expressing hNa_v1.2 and hNa_v1.6 were cultivated in the same conditions mentioned above for the HEK293-hNa_v1.3 cells. For the automated patch-clamp experiments, cells were passaged from 24 to 48 h prior to the experiment and replated in the appropriate quantity to reach 50–70% confluence. For electrophysiological recordings, cells were detached with trypsin-EDTA and diluted in a culture medium. Then, the cells were centrifuged at 800 rpm for 4 min and resuspended (~300,000 cells/mL) in a patch-clamp extracellular solution.

4.4. Fluorescent Assays

Voltage sensor probe (VSP) assays were carried out as described previously [59]. VSP were freshly prepared in an Na-free buffer (VSP Buffer) containing (in mM): 160 TEA-Cl, 0.1 CaCl₂, 1 MgCl₂, 11 Glucose, and 10 HEPES-K (pH 7.4). Cells were washed with the VSP Buffer and first incubated with CC2-DMPE (5 µM) and Pluronic®-F127 acid (0.01%) dissolved in the VSP Buffer for 70 min at RT. After washing with the VSP buffer, the cells were loaded with DisBAC₂(3) (10 µM) and VABSC-1 (250 µM) dissolved and diluted in the VSP Buffer for 45 min at RT. In control experiments, BTX (1–10 µM) with and without TTX (1–10 µM) was added to the VSP buffer containing DisBAC₂(3) and VABSC-1. The plates were illuminated at $\lambda = 405$ nm, and the resulting fluorescence emissions were monitored at $\lambda = 460$ and $\lambda = 580$ nm. The plate reader measured the fluorescence emission every 5 s up to 145 s. After a 30 s baseline, cell depolarization was induced by the injection of a depolarizing solution (in mM: 160 NaCl, 4.5 KCl, 2 CaCl₂, 1 MgCl₂, 11 glucose, 10 HEPES-K, (pH 7.4)), raising the final Na⁺ concentration to 80 mM. The osmolarity of the solution remained the same at around 300 ± 5 mOsmol before and after the addition of the depolarizing solution. For the screening assays, IA were co-incubated at 10–50 µg/mL with

1 μM of BTX in triplicate. In positive control experiments, BTX (1 μM) with TTX (10 μM) and DMSO (1%) were added to the VSP buffer containing DisBAC₂(3) and VABSC-1. To establish the concentration–response relationships, increasing concentrations of compounds were co-incubated with BTX (1 μM). These experiments were performed in three wells and repeated twice. Intracellular Na⁺ fluorescent assays were performed using the cell-permeant acetoxymethyl esters (AM) form of the Asante NaTRIUM Green-2 (ANG-2) probe, a cytosolic Na⁺ indicator. The HEK293-hNav_{1.3} cells were incubated for 1 h in the dark at 37 °C in the ANG-2 probe (5 μM) prepared with Hank’s buffer saline solution (HBSS), which contained (in mM): 2 CaCl₂, 1 MgCl₂, 5 KCl, 150 NaCl, 10 D-glucose, 10 HEPES (pH 7.4), and 0.02% pluronic acid. Afterwards, the quencher solution Ponceau 4R (P4R, 1 mM) was added, and the fluorescence signal was monitored at the following wavelengths: excitation 517 nm and emission 540 nm. Thirty seconds after baseline recording, BTX (1 μM) was automatically injected with or without TTX (1 μM) or IA (10 μM), and the fluorescence emission was read for 1000 s. All experiments were performed in triplicate wells and repeated thrice. Fluorescence measurement was carried out using a FlexStation[®] 3 Benchtop Multi-Mode Microplate Reader (Molecular Devices, Sunnyvale, CA, USA).

4.5. Quantitative Real Time PCR

Total RNA from the GH3b6 cells was extracted using the RNeasy[®] micro kit (Qiagen, Courtaboeuf, France). For retrotranscription, 1 μg of total RNA was used, and the reactions were performed using random hexamer primers and the QuantiTect[®] Reverse Transcription kit (Qiagen). Real-time PCR assays were carried out on a LightCycler[®] 480 Instrument II (Roche, Meylan, France) using the Sybr[®] Select Master Mix (Applied Biosystems) with 2.5, 5, and 10 ng of cDNA in duplicate. Gene-specific primers for real-time PCR (Table 4) were designed using the Primer3 Software. Differences in transcript levels were determined using the cycle threshold method, as described by the manufacturer. Specificity of amplification was confirmed by melting curve analysis. Relative quantification of gene expression was normalized to the mean of expression of two validated House-keeping genes: *gapdh* (glyceraldehyde-3-phosphate dehydrogenase) and *Gusb* (beta-glucuronidase), according to the formula $E = 2^{-(C_t(\text{Target}) - C_{t\text{mean}}(\text{Reference}))}$, where C_t is the threshold cycle. Amplicon sizes (70–106 bp) and AT content (47–55%) were chosen to allow for a comparison of the relative expression values obtained for each gene [60].

Table 4. Primers pairs used to amplify Nav channel transcripts by RTq-PCR.

Gene Name	GenBank Accession Number	Protein Name	Forward Primer (5'–3')	Reverse Primer (5'–3')	Amplicon Size (bp)
<i>scn1a</i>	NM_030875.1	Nav _{1.1}	gttcgacatcgccagtt	catctcagttcagtagttgtcca	92
<i>scn2a</i>	NM_012647.1	Nav _{1.2}	tggtgtccctggttgag	ccttattctgtctcagtagttgtgc	88
<i>scn3a</i>	NM_013119.1	Nav _{1.3}	gcaccgttcattctaaccat	tttagcttctgtcataagaattgc	92
<i>scn4a</i>	NM_013178.1	Nav _{1.4}	ggcactgtctcgatttgagg	ttcatgatggaggggatagc	71
<i>scn5a</i>	NM_013125.2	Nav _{1.5}	tgccaccaatgccttgta	catgatgagcatgctaagagagc	96
<i>scn8a</i>	NM_019266.2	Nav _{1.6}	ggaagttttccatcatgaatcag	gctgttatgtcgggagagga	70
<i>scn9a</i>	NM_133289	Nav _{1.7}	cagcagatgtagaccgactca	actcgtgaactcagcagcag	78
<i>scn10a</i>	NM_017247.1	Nav _{1.8}	agaggacccaagggaca	tggtggtttcacacttttg	59
<i>scn11a</i>	NM_019265.2	Nav _{1.9}	cagaggacgatgcctctaaaa	ttctgggacagtcgtttggt	60
<i>gusb</i>	NM_017015.2	Gus	ctctggtggccttacctgat	cagactcaggtgtgtcatcgc	78
<i>gapdh</i>	NM_017008.3	Gapdh	tggaagctggtcatcaac	gcataccccattgatgtt	77

Primer pairs used to amplify Nav channel transcripts by RTq-PCR. GenBank accession number, sequence, and amplicon size are listed. The primer sequences for the housekeeping gene *gusb* and *gapdh* are also indicated.

4.6. HPLC Analysis

Chromatographic analyses were performed on a 2695 separation module coupled to a 2695 PDA detector (Waters, Saint Quentin en Yvelines, France), assisted by the Empower software (Waters). The IA were dissolved in MeOH at 1 mg/mL and centrifuged for 6 min at 13,000 rpm. The injection volume was 5 μL . Samples were injected into a Lichrospher

100 RP18 column (125 mm × 4 mm, 5 µm, Merck) and were separated by a gradient elution with buffer A (50 mM KH₂PO₄, 2 mM heptane sulfonic acid, 0.1% diethylamine, pH = 3) and the MeCN (B) solvent system (T₀ 5% B; T_{25'} 100% B and T_{30'} 100% B) [61]. The flow rate was 1 mL/min, with UV detection at 210 and 280 nm, respectively.

4.7. Automated Patch-Clamp Experiments

Whole-cell recordings were used to investigate the effects of oxostephanine, liriode-nine, thalamiculine, bebeerine, and protopine on HEK293 cells stably expressing the hNav1.2, hNav1.3, and hNav1.6 channels. Automated patch-clamp recordings were performed using the SyncroPatch 384PE from Nanion (München, Germany). Chips with a single hole (n = 384, series resistance 3.93 ± 1.19 MΩ) were used for HEK293 cell recordings. Pulse generation and data collection were performed with the PatchControl384 v1.9.7 software (Nanion) and the Biomek interface (Beckman Coulter). Whole-cell recordings were conducted according to the recommended procedures of Nanion. The cells were stored in a cell hotel reservoir at 10 °C, with a shaking speed of 60 RPM. After initiating the experiment, cell catch, sealing, whole-cell formation, liquid application, recording, and data acquisition were all performed sequentially and automatically. The intracellular solution contained (in mM): 10 CsCl, 110 CsF, 10 NaCl, 10 EGTA, and 10 HEPES (pH 7.2, osmolarity 280 mOsm), and the extracellular solution contained (in mM): 60 NMDG, 80 NaCl, 4 KCl, 2 CaCl₂, 1 MgCl₂, 5 Glucose, and 10 HEPES (pH 7.4 (NaOH), osmolarity 280 ± 3 mOsm). Whole-cell experiments were performed at a holding potential of −100 mV at room temperature (18–22 °C). Currents were sampled at 20 kHz. For pharmacological assays, alkaloid solutions were prepared at 30 µM in the extracellular solution supplemented with 0.3% bovine serum albumin and distributed in 384-well compound plates according to a pre-established plate plan. This distribution within compound plates could not be randomized since it would then excessively complexify the methods of analyses. The working compound solution was diluted 3 times in the patch-clamp recording well by adding from 30 to 60 µL of the external solution to reach the final reported concentration and the test volume of 90 µL. The effects of IA (10 µM) or of BTX (1 µM) were measured at the end of a 10 min application time. Activation curves were built by depolarizations lasting for 2000 ms, from −130 mV to 40 mV, using 5 mV increment steps. Pulses were applied every 5 s. The use dependency of oxostephanine and the liriode-nine effects on hNav1.3 with or without the presence of 1 µM BTX were investigated by depolarizing the cells to 10 mV at 1 Hz for 300 s.

4.8. Structural Analysis

¹H NMR spectra were conducted in the appropriate deuterated solvent (CHCl₃-d or CH₃OH-d₄) on a Bruker Avance DRX-500 spectrometer (Bruker France, Wissembourg, France) operating at 500 MHz (¹H). Fast Atom Bombardment (FAB) mass spectra were obtained on a B/E JMS 700 MStation spectrometer (Jeol Europe, Croissy-sur-Seine, France), with 3-nitrobenzyl alcohol as the matrix and argon as the collision gas. Optical rotations ($[\alpha]_D^{T^\circ C}$) were measured at 20 °C on a Polartronic I polarimeter (Schmidt-Haensch, Berlin, Germany).

4.9. Data Bank Search

Structural analogs of oxostephanine and liriode-nine were searched for using the search tool from ChemIDplus (<https://chem.nlm.nih.gov/chemidplus/ProxyServlet>, June 2021). Analogs sharing less than 94% were ignored.

4.10. Data Analysis

Fluorescence data analysis was performed using the SoftMax Pro 5.4.1 software (Molecular Devices, Sunnyvale, CA, USA).

With data from VSP assays, calculations made for histograms were performed as follows. For t = 1 s up to 30 s, for each condition, the VSP signal ratio of the GH3b6 cells

was measured after 30 min of incubation with 1 μ M BTX and an individual alkaloid. The first 30 s showed signal at equilibrium before the addition of the depolarizing solution, resulting in a final Na^+ concentration of 80 mM. Each curve point is the CC2DMPE signal over the DisBAC(2,3) ratio, as a ratio relative to the response before the addition of the depolarizing solution (calculated by averaging the first 4 time points, T1-4, from each replicate). The baseline of this ratio was shifted to 0 by subtracting the average of the 4 first time points before adding the depolarizing solution. Each value was then expressed as a percentage of the positive control final ratio obtained in each plate/experiment with 1 μ M BTX. This allowed us to compare data from different microplates. The equation used was the following:

$$\frac{\left(\frac{\text{CC2DMPE/DisBAC2,3}}{\text{Average}_{\text{T1-4}}(\text{CC2DMPE/DisBAC2,3})} - 1 \right) \times 100}{\left(\frac{\text{AverageControlBTX}_{\text{T27-30}}(\text{CC2DMPE/DisBAC2,3})}{\text{AverageControlBTX}_{\text{T1-4}}(\text{CC2DMPE/DisBAC2,3})} - 1 \right)}$$

These values were plotted over time (Figure 4a). The histograms (Figure 4b) represent the inhibitory effect of each IA, normalized as follows: 100% (VSP signal induced by BTX at 1 μ M) average of the last 4 values of each curve.

The kinetic traces of the ANG-2 fluorescence were plotted as an emission ratio: $\lambda_{\text{excitation 517 nm}}/\lambda_{\text{excitation 540 nm}}$. The AUC of the kinetic traces were used to determine the response after BTX injection, alone (1 μ M) or with each IA (10 μ M).

All graphs and statistical analyses were completed using the GraphPad Prism 7.02 (La Jolla, CA, USA). Data are presented as mean $\pm$ SEM, calculated from 3 replicates and representative of 3 independent experiments. Non-linear analysis was used to fit the concentration–response data with the Langmuir–Hill equation with a variable slope. Multiple groups were compared by using a one-way analysis of variance (ANOVA), followed by a Bonferroni post hoc test, when appropriate. Differences between independent groups were assessed by using parametric or non-parametric *t*-tests (Mann and Whitney) when appropriate. Differences with $p < 0.05$ were considered significant (* for $p < 0.05$, ** for $p < 0.01$, *** for $p < 0.001$, **** for $p < 0.0001$).

5. Conclusions

In conclusion, the VSP screening of the in-house alkaloid library allowed for the identification of five IA—oxostephanine, liriodenine, thalamiculine, bebeerine, and protopine—as inhibitors of BTX-activated Na_v channels. However, only bebeerine was validated as a potent blocker of Na_v channels by automated patch-clamp experiments, with the following rank order: $\text{hNa}_v1.6 > \text{hNa}_v1.3 > \text{hNa}_v1.2$. In fact, oxostephanine and liriodenine were able to decrease the BTX-induced alteration of both the activation and inactivation properties of the $\text{hNa}_v1.3$ channel. Our data highlight that the use of the Na_v channel activator, which binds within the pore, could lead to the selection of low-potency Na_v channel inhibitors, and that electrophysiology is unavoidable for their characterization. Finally, bebeerine is the first BBIQ exhibiting potent inhibitory effects on the $\text{hNa}_v1.3$ and $\text{hNa}_v1.6$ channels.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/molecules27134133/s1>, Figure S1: ^1H NMR spectrum of IA24 and IA14 (oxostephanine); Figure S2: ^1H NMR spectrum of IA39 (liriodenine); Figure S3: ^1H NMR spectrum of IA49 (thalamiculine); Figure S4: ^1H NMR spectrum of IA52 (bebeerine); Figure S5: ^1H NMR spectrum of IA69 (protopine); Figure S6: World map showing the geographical origins of plants used to purify the 62 isoquinoline alkaloids screened for voltage-gated Na^+ channel blockers; Figure S7: Effects of oxostephanine, liriodenine, thalamiculine, bebeerine and protopine on Na^+ currents elicited by $\text{hNa}_v1.2$, $\text{hNa}_v1.3$ and $\text{hNa}_v1.6$ channels; Table S1: Alkaloids used in this work to screen for Na_v channel inhibitors.

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Sample Availability: Samples of the compounds are available from the authors.

Abbreviations

The following abbreviations are used in this manuscript:

AUC	area under the curve
BTX	batrachotoxin
BBIQ	bisbenzylisoquinoline
IA	isoquinoline alkaloids
[Na ⁺] _i	intracellular Na ⁺ concentration
Nav channel	voltage-gated Na ⁺ channel
TTX	tetrodotoxin
TTX-R	resistant to tetrodotoxin
TTX-S	sensitive to tetrodotoxin
VTD	veratridine

References

- Hille, B. *Ion Channels of Excitable Membranes*, 3rd ed. Sinauer: Sunderland, MA, USA, 2001; ISBN 978-0-87893-321-1.
- Catterall, W.A.; Goldin, A.L.; Waxman, S.G. International Union of Pharmacology. XLVII. Nomenclature and Structure-Function Relationships of Voltage-Gated Sodium Channels. *Pharmacol. Rev.* **2005**, *57*, 397–409. [\[CrossRef\]](#) [\[PubMed\]](#)
- Eijkelkamp, N.; Linley, J.E.; Baker, M.D.; Minett, M.S.; Clegg, R.; Werdehausen, R.; Rugiero, F.; Wood, J.N. Neurological Perspectives on Voltage-Gated Sodium Channels. *Brain* **2012**, *135*, 2585–2612. [\[CrossRef\]](#) [\[PubMed\]](#)
- De Lera Ruiz, M.; Kraus, R.L. Voltage-Gated Sodium Channels: Structure, Function, Pharmacology, and Clinical Indications. *J. Med. Chem.* **2015**, *58*, 7093–7118. [\[CrossRef\]](#) [\[PubMed\]](#)
- Azimova, S.S.; Yunusov, M.S. (Eds.) *Natural Compounds: Plant Sources, Structure and Properties*; Springer reference; Springer: New York, NY, USA, 2013; ISBN 978-1-4614-0542-9.
- Amirkia, V.; Heinrich, M. Alkaloids as Drug Leads – A Predictive Structural and Biodiversity-Based Analysis. *Phytochem. Lett.* **2014**, *10*, xlviii–liii. [\[CrossRef\]](#)
- Wink, M. Modes of Action of Herbal Medicines and Plant Secondary Metabolites. *Medicines* **2015**, *2*, 251–286. [\[CrossRef\]](#)
- Wang, S.Y.; Wang, G.K. Voltage-Gated Sodium Channels as Primary Targets of Diverse Lipid-Soluble Neurotoxins. *Cell. Signal.* **2003**, *15*, 151–159. [\[CrossRef\]](#)
- Kontis, K.J.; Goldin, A.L. Site-Directed Mutagenesis of the Putative Pore Region of the Rat IIA Sodium Channel. *Mol. Pharmacol.* **1993**, *43*, 635–644. [\[PubMed\]](#)
- Terlau, H.; Heinemann, S.H.; Stuhmer, W.; Pusch, M.; Conti, F.; Imoto, K.; Numa, S. Mapping the Site of Block by Tetrodotoxin and Saxitoxin of Sodium Channel II. *FEBS Lett.* **1991**, *293*, 93–96. [\[CrossRef\]](#)
- Durán-Riveroll, L.; Cembella, A. Guanidinium Toxins and Their Interactions with Voltage-Gated Sodium Ion Channels. *Mar. Drugs* **2017**, *15*, 303. [\[CrossRef\]](#) [\[PubMed\]](#)

12. Tsukamoto, T.; Chiba, Y.; Wakamori, M.; Yamada, T.; Tsunogae, S.; Cho, Y.; Sakakibara, R.; Imazu, T.; Tokoro, S.; Satake, Y.; et al. Differential Binding of Tetrodotoxin and Its Derivatives to Voltage-sensitive Sodium Channel Subtypes (Na_v 1.1 to Na_v 1.7). *J. Cereb. Blood Flow Metab.* **2017**, *174*, 3881–3892. [[CrossRef](#)] [[PubMed](#)]
13. Catterall, W.A. Activation of the Action Potential Na⁺ Ionophore by Neurotoxins. An Allosteric Model. *J. Biol. Chem.* **1977**, *252*, 8669–8676. [[CrossRef](#)]
14. Tamkun, M.M.; Catterall, W.A. Ion Flux Studies of Voltage-Sensitive Sodium Channels in Synaptic Nerve-Ending Particles. *Mol. Pharmacol.* **1981**, *19*, 78–86. [[PubMed](#)]
15. Ameri, A.; Simmet, T. Antagonism of the Aconitine-Induced Inexcitability by the Structurally Related Aconitum Alkaloids, Lappaconitine and Ajacine. *Brain Res.* **1999**, *842*, 332–341. [[CrossRef](#)]
16. Li, Y.; Zheng, Y.; Yu, Y.; Gan, Y.; Gao, Z. Inhibitory Effects of Lappaconitine on the Neuronal Isoforms of Voltage-Gated Sodium Channels. *Acta Pharmacol. Sin.* **2019**, *40*, 451–459. [[CrossRef](#)]
17. Sun, M.-L.; Ao, J.-P.; Wang, Y.-R.; Huang, Q.; Li, T.-F.; Li, X.-Y.; Wang, Y.-X. Lappaconitine, a C18-Diterpenoid Alkaloid, Exhibits Antihypersensitivity in Chronic Pain through Stimulation of Spinal Dynorphin A Expression. *Psychopharmacology* **2018**, *235*, 2559–2571. [[CrossRef](#)] [[PubMed](#)]
18. Fischer, F.; Vonderlin, N.; Zitron, E.; Seyler, C.; Scherer, D.; Becker, R.; Katus, H.A.; Scholz, E.P. Inhibition of Cardiac Kv1.5 and Kv4.3 Potassium Channels by the Class Ia Anti-Arrhythmic Ajmaline: Mode of Action. *Naunyn-Schmiedeberg's Arch. Exp. Pathol. Pharmacol.* **2013**, *386*, 991–999. [[CrossRef](#)] [[PubMed](#)]
19. Friedrich, O.; V Wegner, F.; Wink, M.; Fink, R.H.A. Na⁺ and K⁺ -Channels as Molecular Targets of the Alkaloid Ajmaline in Skeletal Muscle Fibres: Ajmaline Action on Skeletal Muscle Cation Channels. *J. Cereb. Blood Flow Metab.* **2007**, *151*, 63–74. [[CrossRef](#)] [[PubMed](#)]
20. Kiesecker, C.; Zitron, E.; Luck, S.; Bloehs, R.; Scholz, E.P.; Kathofer, S.; Thomas, D.; Kreye, V.A.W.; Katus, H.A.; Schoels, W.; et al. Class Ia Anti-Arrhythmic Drug Ajmaline Blocks HERG Potassium Channels: Mode of Action. *Naunyn-Schmiedeberg's Arch. Exp. Pathol. Pharmacol.* **2004**, *370*, 423–435. [[CrossRef](#)] [[PubMed](#)]
21. Miller, D.C.; Harmer, S.C.; Poliandri, A.; Nobles, M.; Edwards, E.C.; Ware, J.S.; Sharp, T.V.; McKay, T.R.; Dunkel, L.; Lambiase, P.D.; et al. Ajmaline Blocks I_{Na} and I_{Kr} without Eliciting Differences between Brugada Syndrome Patient and Control Human Pluripotent Stem Cell-Derived Cardiac Clusters. *Stem Cell Res.* **2017**, *25*, 233–244. [[CrossRef](#)]
22. Rolf, S.; Bruns, H.; Wichter, T.; Kirchhof, P.; Ribbing, M.; Wasmer, K.; Paul, M.; Breithardt, G.; Haverkamp, W.; Eckardt, L. The Ajmaline Challenge in Brugada Syndrome: Diagnostic Impact, Safety, and Recommended Protocol. *Eur. Hear. J.* **2003**, *24*, 1104–1112. [[CrossRef](#)]
23. Yu, G.; Qian, L.; Yu, J.; Tang, M.; Wang, C.; Zhou, Y.; Geng, X.; Zhu, C.; Yang, Y.; Pan, Y.; et al. Brucine Alleviates Neuropathic Pain in Mice via Reducing the Current of the Sodium Channel. *J. Ethnopharmacol.* **2019**, *233*, 56–63. [[CrossRef](#)] [[PubMed](#)]
24. Splettstoesser, F.; Bonnet, U.; Wiemann, M.; Bingmann, D.; Büsselberg, D. Modulation of Voltage-Gated Channel Currents by Harmaline and Harmane: Harmala Alkaloids Modulate Voltage-Gated Currents. *J. Cereb. Blood Flow Metab.* **2005**, *144*, 52–58. [[CrossRef](#)] [[PubMed](#)]
25. Zhao, F.; Tang, Q.; Xu, J.; Wang, S.; Li, S.; Zou, X.; Cao, Z. Dehydrocrenatidine Inhibits Voltage-Gated Sodium Channels and Ameliorates Mechanic Allodia in a Rat Model of Neuropathic Pain. *Toxins* **2019**, *11*, 229. [[CrossRef](#)] [[PubMed](#)]
26. Bentley, K.W. β -Phenylethylamines and the Isoquinoline Alkaloids. *Nat. Prod. Rep.* **2006**, *23*, 444–463. [[CrossRef](#)] [[PubMed](#)]
27. Chang, G.-J.; Wu, M.-H.; Wu, Y.-C.; Su, M.-J. Electrophysiological Mechanisms for Antiarrhythmic Efficacy and Positive Inotropy of Liriodenine, a Natural Aporphine Alkaloid from *Fissistigma Glaucescens*. *J. Cereb. Blood Flow Metab.* **1996**, *118*, 1571–1583. [[CrossRef](#)]
28. Xiao-Shan, H.; Qing, L.; Yun-Shu, M.; Ze-Pu, Y. Crebanine Inhibits Voltage-Dependent Na⁺ Current in Guinea-Pig Ventricular Myocytes. *Chin. J. Nat. Med.* **2014**, *12*, 20–23. [[CrossRef](#)]
29. Wang, J.; Ma, Y.; Sachs, F.; Li, J.; Suchyna, T.M. GsMTx4-D Is a Cardioprotectant against Myocardial Infarction during Ischemia and Reperfusion. *J. Mol. Cell. Cardiol.* **2016**, *98*, 83–94. [[CrossRef](#)]
30. Araújo, D.A.M.; Mafra, R.A.; Rodrigues, A.L.P.; Miguel-Silva, V.; Beirão, P.S.L.; De Almeida, R.N.; Quintans, L.; Souza, M.F.V.de; Cruz, J.S. N-Salicyloyltryptamine, a New Anticonvulsant Drug, Acts on Voltage-Dependent Na⁺, Ca²⁺, and K⁺ Ion Channels: N-Salicyloyltryptamine Acts on Voltage-Dependent Ion Channels. *J. Cereb. Blood Flow Metab.* **2003**, *140*, 1331–1339. [[CrossRef](#)]
31. Baroni, D.; Picco, C.; Barbieri, R.; Moran, O. Antisense-Mediated Post-Transcriptional Silencing of *SCN1B* Gene Modulates Sodium Channel Functional Expression: Silencing of *SCN1B* Gene Modulates the Sodium Channel Expression. *Biol. Cell* **2014**, *106*, 13–29. [[CrossRef](#)]
32. Campos, F.V.; Coronas, F.I.V.; Beirão, P.S.L. Voltage-Dependent Displacement of the Scorpion Toxin Ts3 from Sodium Channels and Its Implication on the Control of Inactivation: Scorpion Toxin and Sodium Channel Inactivation. *J. Cereb. Blood Flow Metab.* **2004**, *142*, 1115–1122. [[CrossRef](#)]
33. So, E.C.; Wu, S.-N.; Lo, Y.-C.; Su, K. Differential Regulation of Tefluthrin and Telmisartan on the Gating Charges of I_{Na} Activation and Inactivation as Well as on Resurgent and Persistent I_{Na} in a Pituitary Cell Line (GH3). *Toxicol. Lett.* **2018**, *285*, 104–112. [[CrossRef](#)] [[PubMed](#)]
34. Wu, S.-N.; Wu, Y.-H.; Chen, B.-S.; Lo, Y.-C.; Liu, Y.-C. Underlying Mechanism of Actions of Tefluthrin, a Pyrethroid Insecticide, on Voltage-Gated Ion Currents and on Action Currents in Pituitary Tumor (GH3) Cells and GnRH-Secreting (GT1-7) Neurons. *Toxicology* **2009**, *258*, 70–77. [[CrossRef](#)] [[PubMed](#)]

35. Réthoré, L.; Park, J.; Montnach, J.; Nicolas, S.; Khoury, J.; Le Seac'h, E.; Mabrouk, K.; De Pomyers, H.; Tricoire-Leignel, H.; Mattei, C.; et al. Pharmacological Dissection of the Crosstalk between NaV and CaV Channels in GH3b6 Cells. *Int. J. Mol. Sci.* **2022**, *23*, 827. [\[CrossRef\]](#) [\[PubMed\]](#)
36. Baroni, D.; Moran, O. Molecular Differential Expression of Voltage-Gated Sodium Channel Alpha and Beta Subunit MRNAs in Five Different Mammalian Cell Lines. *J. Bioenerg. Biomembr.* **2011**, *43*, 729–738. [\[CrossRef\]](#)
37. Vega, A.V.; Espinosa, J.L.; Lopez-Dominguez, A.M.; Lopez-Santiago, L.F.; Navarrete, A.; Cota, G. L-Type Calcium Channel Activation up-Regulates the MRNAs for Two Different Sodium Channel Alpha Subunits (Nav1.2 and Nav1.3) in Rat Pituitary GH3 Cells. *Mol. Brain Res.* **2003**, *116*, 115–125. [\[CrossRef\]](#)
38. Iamshanova, O.; Mariot, P.; Lehen'kyi, V.; Prevarskaya, N. Comparison of Fluorescence Probes for Intracellular Sodium Imaging in Prostate Cancer Cell Lines. *Eur. Biophys. J.* **2016**, *45*, 765–777. [\[CrossRef\]](#)
39. Stojilkovic, S.S.; Tabak, J.; Bertram, R. Ion Channels and Signaling in the Pituitary Gland. *Endocr. Rev.* **2010**, *31*, 845–915. [\[CrossRef\]](#)
40. Yang, L.; Li, Q.; Liu, X.; Liu, S. Roles of Voltage-Gated Tetrodotoxin-Sensitive Sodium Channels Nav1.3 and Nav1.7 in Diabetes and Painful Diabetic Neuropathy. *Int. J. Mol. Sci.* **2016**, *17*, 1479. [\[CrossRef\]](#)
41. Zhao, F.; Li, X.; Jin, L.; Zhang, F.; Inoue, M.; Yu, B.; Cao, Z. Development of a Rapid Throughput Assay for Identification of HNav1.7 Antagonist Using Unique Efficacious Sodium Channel Agonist, Antillatoxin. *Mar. Drugs.* **2016**, *14*, 36. [\[CrossRef\]](#)
42. Gonzalez, J.E.; Tsien, R.Y. Improved Indicators of Cell Membrane Potential That Use Fluorescence Resonance Energy Transfer. *Chem. Biol.* **1997**, *4*, 269–277. [\[CrossRef\]](#)
43. Liu, C.J.; Priest, B.T.; Bugianesi, R.M.; Dulski, P.M.; Felix, J.P.; Dick, I.E.; Brochu, R.M.; Knaus, H.G.; Middleton, R.E.; Kaczorowski, G.J.; et al. A High-Capacity Membrane Potential FRET-Based Assay for Nav1.8 Channels. *Assay Drug Dev. Technol.* **2006**, *4*, 37–48. [\[CrossRef\]](#)
44. Middleton, R.E.; Warren, V.A.; Kraus, R.L.; Hwang, J.C.; Liu, C.J.; Dai, G.; Brochu, R.M.; Kohler, M.G.; Gao, Y.D.; Garsky, V.M.; et al. Two Tarantula Peptides Inhibit Activation of Multiple Sodium Channels. *Biochemistry* **2002**, *41*, 14734–14747. [\[CrossRef\]](#) [\[PubMed\]](#)
45. Williams, B.S.; Felix, J.P.; Priest, B.T.; Brochu, R.M.; Dai, K.; Hoyt, S.B.; London, C.; Tang, Y.S.; Duffy, J.L.; Parsons, W.H.; et al. Characterization of a New Class of Potent Inhibitors of the Voltage-Gated Sodium Channel Nav1.7. *Biochemistry* **2007**, *46*, 14693–14703. [\[CrossRef\]](#) [\[PubMed\]](#)
46. Deuis, J.R.; Mueller, A.; Israel, M.R.; Vetter, I. The Pharmacology of Voltage-Gated Sodium Channel Activators. *Neuropharmacology* **2017**, *127*, 87–108. [\[CrossRef\]](#)
47. Chernov-Rogan, T.; Li, T.; Lu, G.; Verschoof, H.; Khakh, K.; Jones, S.W.; Beresini, M.H.; Liu, C.; Ortwine, D.F.; McKerrall, S.J.; et al. Mechanism-Specific Assay Design Facilitates the Discovery of Nav1.7-Selective Inhibitors. *Proc. Natl. Acad. Sci. USA* **2018**, *115*, E792–E801. [\[CrossRef\]](#)
48. Felix, J.P.; Williams, B.S.; Priest, B.T.; Brochu, R.M.; Dick, I.E.; Warren, V.A.; Yan, L.; Slaughter, R.S.; Kaczorowski, G.J.; Smith, M.M.; et al. Functional Assay of Voltage-Gated Sodium Channels Using Membrane Potential-Sensitive Dyes. *Assay Drug Dev. Technol.* **2004**, *2*, 260–268. [\[CrossRef\]](#)
49. Du, Y.; Days, E.; Romaine, I.; Abney, K.K.; Kaufmann, K.; Sulikowski, G.; Stauffer, S.; Lindsley, C.W.; Weaver, C.D. Development and Validation of a Thallium Flux-Based Functional Assay for the Sodium Channel Nav1.7 and Its Utility for Lead Discovery and Compound Profiling. *ACS Chem. Neurosci.* **2015**, *6*, 871–878. [\[CrossRef\]](#) [\[PubMed\]](#)
50. Du, Y.; Garden, D.P.; Wang, L.; Zhorov, B.S.; Dong, K. Identification of New Batrachotoxin-Sensing Residues in Segment IIIS6 of the Sodium Channel. *J. Biol. Chem.* **2011**, *286*, 13151–13160. [\[CrossRef\]](#) [\[PubMed\]](#)
51. Chang, K.C.; Su, M.J.; Peng, Y.I.; Shao, C.C.; Wu, Y.C.; Tseng, Y.Z. Mechanical Effects of Liriodenine on the Left Ventricular-Arterial Coupling in Wistar Rats: Pressure-Stroke Volume Analysis. *Br. J. Pharmacol.* **2001**, *133*, 29–36. [\[CrossRef\]](#)
52. Khamis, S.; Bibby, M.C.; Brown, J.E.; Cooper, P.A.; Scowen, I.; Wright, C.W. Phytochemistry and Preliminary Biological Evaluation of Cyathostemma Argenteum, a Malaysian Plant Used Traditionally for the Treatment of Breast Cancer. *Phytother. Res.* **2004**, *18*, 507–510. [\[CrossRef\]](#)
53. Wang, T.S.; Luo, Y.P.; Wang, J.; He, M.X.; Zhong, M.G.; Li, Y.; Song, X.P. (+)-Rumphiin and Polyalthurea, New Compounds from the Stems of Polyalthia Rumphii. *Nat. Prod. Commun.* **2013**, *8*, 1427–1429. [\[CrossRef\]](#)
54. Chen, C. Review on Pharmacological Activities of Liriodenine. *Afr. J. Pharm. Pharmacol.* **2013**, *7*, 1067–1070. [\[CrossRef\]](#)
55. Coquerel, Q.R.R.; Demares, F.; Geldenhuys, W.J.; Le Ray, A.M.; Breard, D.; Richomme, P.; Legros, C.; Norris, E.; Bloomquist, J.R. Toxicity and Mode of Action of the Aporphine Plant Alkaloid Liriodenine on the Insect GABA Receptor. *Toxicon* **2021**, *201*, 141–147. [\[CrossRef\]](#) [\[PubMed\]](#)
56. Lin, C.H.; Yang, C.M.; Ko, F.N.; Wu, Y.C.; Teng, C.M. Antimuscarinic Action of Liriodenine, Isolated from Fissistigma Glaucescens, in Canine Tracheal Smooth Muscle. *Br. J. Pharmacol.* **1994**, *113*, 1464–1470. [\[CrossRef\]](#) [\[PubMed\]](#)
57. Medeiros, M.A.A.; Pinho, J.F.; De-Lira, D.P.; Barbosa-Filho, J.M.; Araújo, D.A.M.; Cortes, S.F.; Lemos, V.S.; Cruz, J.S. Curine, a Bisbenzylisoquinoline Alkaloid, Blocks L-Type Ca²⁺ Channels and Decreases Intracellular Ca²⁺ Transients in A7r5 Cells. *Eur. J. Pharmacol.* **2011**, *669*, 100–107. [\[CrossRef\]](#) [\[PubMed\]](#)
58. Tashjian, A.H. Clonal Strains of Hormone-Producing Pituitary Cells. In *Methods in Enzymology*; Elsevier: Amsterdam, The Netherlands, 1979; Volume 58, pp. 527–535. ISBN 978-0-12-181958-3.
59. Priest, B.T.; Garcia, M.L.; Middleton, R.E.; Brochu, R.M.; Clark, S.; Dai, G.; Dick, I.E.; Felix, J.P.; Liu, C.J.; Reisetter, B.S.; et al. A Disubstituted Succinamide Is a Potent Sodium Channel Blocker with Efficacy in a Rat Pain Model. *Biochemistry* **2004**, *43*, 9866–9876. [\[CrossRef\]](#)

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60. Colborn, J.M.; Kosoy, M.Y.; Motin, V.L.; Telepnev, M.V.; Valbuena, G.; Myint, K.S.; Fofanov, Y.; Putonti, C.; Feng, C.; Peruski, L. Improved Detection of Bartonella DNA in Mammalian Hosts and Arthropod Vectors by Real-Time PCR Using the NADH Dehydrogenase Gamma Subunit (NuoG). *J. Clin. Microbiol.* **2010**, *48*, 4630–4633. [[CrossRef](#)] [[PubMed](#)]
 61. Sun, J.; Sun, B.; Han, F.; Yan, S.; Yang, H.; Akio, K. Rapid HPLC Method for Determination of 12 Isoflavone Components in Soybean Seeds. *Agric. Sci. China* **2011**, *10*, 70–77. [[CrossRef](#)]